

REVIEW

Metabolic link between obesity and autoimmune diseases

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ABSTRACT. The abnormal accumulation of visceral adipose tissue in obesity is associated with metabolic changes that include altered glucose tolerance, insulin resistance, hyperlipidemia, and metabolic syndrome. Obesity also coincides with increased incidence of autoimmune diseases. Accumulating evidence suggest that prolonged metabolic overload related to overnutrition, influenced by genetic and epigenetic factors, might affect immunologic self-tolerance through changes in the energy metabolism of immune cells, particularly regulatory T (Treg) cells. A strong activation of nutrient-energy signaling pathways blocks the induction of the transcription factor forkhead P3 (FOXP3), a master regulator of Treg cells, consequently inhibiting their generation and proliferation, thereby promoting proinflammatory response. Expanding our knowledge on the topic, particularly on metabolic T cell flexibility *in vivo* will provide new insights that can be used to develop therapeutic strategies for various inflammatory diseases, including obesity and autoimmune diseases. Targeting specific metabolic pathways is emerging as an important approach to control immune response and maintain immunological homeostasis.

Key words: obesity, autoimmunity, immunometabolism, lymphocyte metabolism, cytokines

Autoimmune diseases, such as many other complex disorders, are believed to arise from a combination of genetic and environmental factors. The most common hypothesis on etiology of autoimmune diseases states that polymorphisms in various genes that are mostly involved in immunity result in defective regulation or reduced threshold for lymphocyte activation, and environmental factors initiate or augment activation of self-reactive lymphocytes that have escaped central thymic control and are poised to react against self-antigens. Although the genetic origins, environmental factors, and clinical manifestations of autoimmunity are vast, many autoimmune diseases share common features that contribute to pathogenesis, such as activation and generation of pathogenic effector CD4⁺ and CD8⁺ T cells. The consensus is that various cytokines present in the microenvironment such as interleukin-1 β (IL-1 β), IL-6, IL-23, and tumor growth factor- β directly affect T cell differentiation into pathogenic effector cells, by promoting T helper 17 (Th17)-cell differentiation while blocking regulatory T (Treg)-cell differentiation [1]. Effector T cells are highly dependent on macronutrients, such as carbohydrates, proteins, and lipids, which derive from their microenvironment, and limited nutrient availability can constrain effector T cell response and promote a tolerogenic environment [2]. Availability of metabo-

lites therefore affects metabolic pathway usage and differentiation of effector T cells, reciprocally influencing the microenvironment.

Autoimmune diseases affect approximately 5% to 9.4% of the EU and US populations, more frequently women than man, and often congregate in families, suggesting genetic predispositions [3, 4]. Their incidence and prevalence between 1985 and 2005 showed a net percent increase per year in incidence ($19.1 \pm 43.1\%$) and prevalence ($12.5 \pm 7.9\%$) [3, 5]. The proposed explanation for recent rapid increase in incidence and prevalence of autoimmune diseases is the change of various environmental factors, including the decrease in infectious diseases due to improved hygienic standards, increased exposure to environmental pollutants and low vitamin D intake, among others [6, 7]. The list of environmental factors also includes an increased daily caloric intake of processed foods [8]. Since the 1970s, there has been an average increase of approximately 35% in the availability of calories per capita, in kilocalories per day, which corresponds to about 2800 kilocalories per day [9]. Industrialized or western food has often a high content of the so-called “holy triad”, namely sugar, fat, and salt, which hyperactivate intracellular nutrient-energy-sensing pathways [10]. For obvious reasons, prolonged overnutrition has also been linked to the dramatic

increase in prevalence of obesity, although other factors, both genetic and environmental, contribute to this global health problem. In 2016, more than 1.9 billion adults, representing 39% of the world's adult population, were affected by overweight, of whom over 650 million (13%) had obesity, with obesity rates surpassing 50% in many countries [11]. A high number of children and adolescents are also affected: in 2016, some 41 million children under the age of 5 years and 340 million (18%) of those aged 5 to 19 years had developed overweight or obesity.

Numerous epidemiologic studies have shown the association between an elevated body mass index and a waist-to-hip ratio, the commonly used anthropometric estimates of fatness, and autoimmune diseases, including type 1 diabetes (T1D), multiple sclerosis (MS), psoriasis, psoriatic arthritis, rheumatoid arthritis (RA), systemic lupus erythematosus (SLE), inflammatory bowel diseases, etc. [12-17]. Excessive adiposity is frequently accompanied by a low-grade chronic inflammatory state marked out by increased systemic markers of inflammation [18]. Although the initial trigger for this inflammation is not fully understood, it is reportedly associated to a homeostatic stress caused by a positive energy balance. Under the prolonged overnutrition conditions, the sensors on the immune cells can directly recognize increased nutrient availability, resulting in their aberrant activation [19]. The locally induced inflammation characterized by elevated secretion of proinflammatory cytokines and adipokines produced by adipose tissue promote immune responses by Th1 and Th17 cells, decrease the level of Treg cells and enhance M1-macrophage polarization [20] (*figure 1*). This proinflammatory microenvironment in turn favors bystander activation of other immune cells, including lymphocytes with autoreactive potential [21]. Obesity-induced adipose-tissue inflammation is associated with broad metabolic changes that include altered glucose tolerance, insulin resistance, hyperlipidemia, and metabolic syndrome. Similar metabolic alterations at a systemic and/or local level are also observed in

autoimmune diseases [22]. This review is aimed to explore the association between obesity and autoimmunity, focusing particularly on the emerging metabolic link.

OBEesity-ASSOCIATED ALTERATIONS IN T CELL METABOLISM AND FUNCTION

The discovery of intracellular nutrients and energy sensors that react on changes in both the extracellular and intracellular environment has shed new light on association between metabolism and T cell differentiation and function. Crucial pathways involved in sensing the presence of nutrients necessary for the generation of energy and maintenance of cellular processes include mechanistic target of rapamycin (mTOR), adenosine monophosphate-activated protein kinase (AMPK), and general-controlled nonrepressed kinase [23, 24]. They sense not only essential nutrients, such as glucose, amino acids, and lipids, but also growth factors linked to nutrition. By controlling the metabolic switching among specific intracellular processes, such as glycolysis, fatty acid oxidation, and oxidative phosphorylation, they have a critical impact on metabolism, immune responses, and inflammation both at the intracellular and at the systemic level [23, 24]. mTOR is involved in the control of anabolic pathways, glycolysis, and nucleotide synthesis. It also suppresses the key catabolic process of autophagy and maintains homeostasis in response to changing availability of nutrients, growth factors (e.g., insulin), and adipocytokines, such as leptin [24]. It has been shown that propagation and integration of T-cell stimulation and costimulation critically depends on the phosphoinositide 3-kinase (PI3K)-AKT-mTOR signaling axis [25]. Naive CD4 activated T cells can differentiate into multiple effector Th-cell lineages, including Th1, Th2, Th17, and T-follicular helper cells, whereas CD8⁺ T cells primarily differentiate into cytotoxic T lymphocytes (CTLs) (*figure 2*). Activation of the PI3K-AKT-mTOR signaling pathway is essential for the generation and metabolic reprogramming of

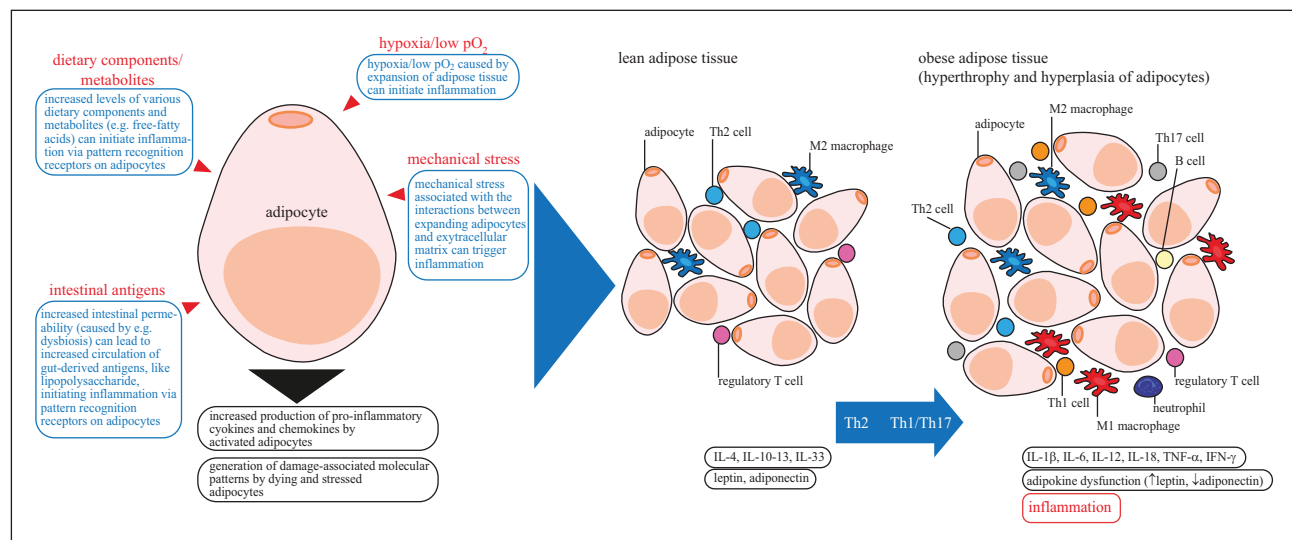


Figure 1

Chronic low-grade systemic inflammation in obesity [20].

IL: interleukin; IFN: interferon; Th: T helper; TFN: tumor necrosis factor.

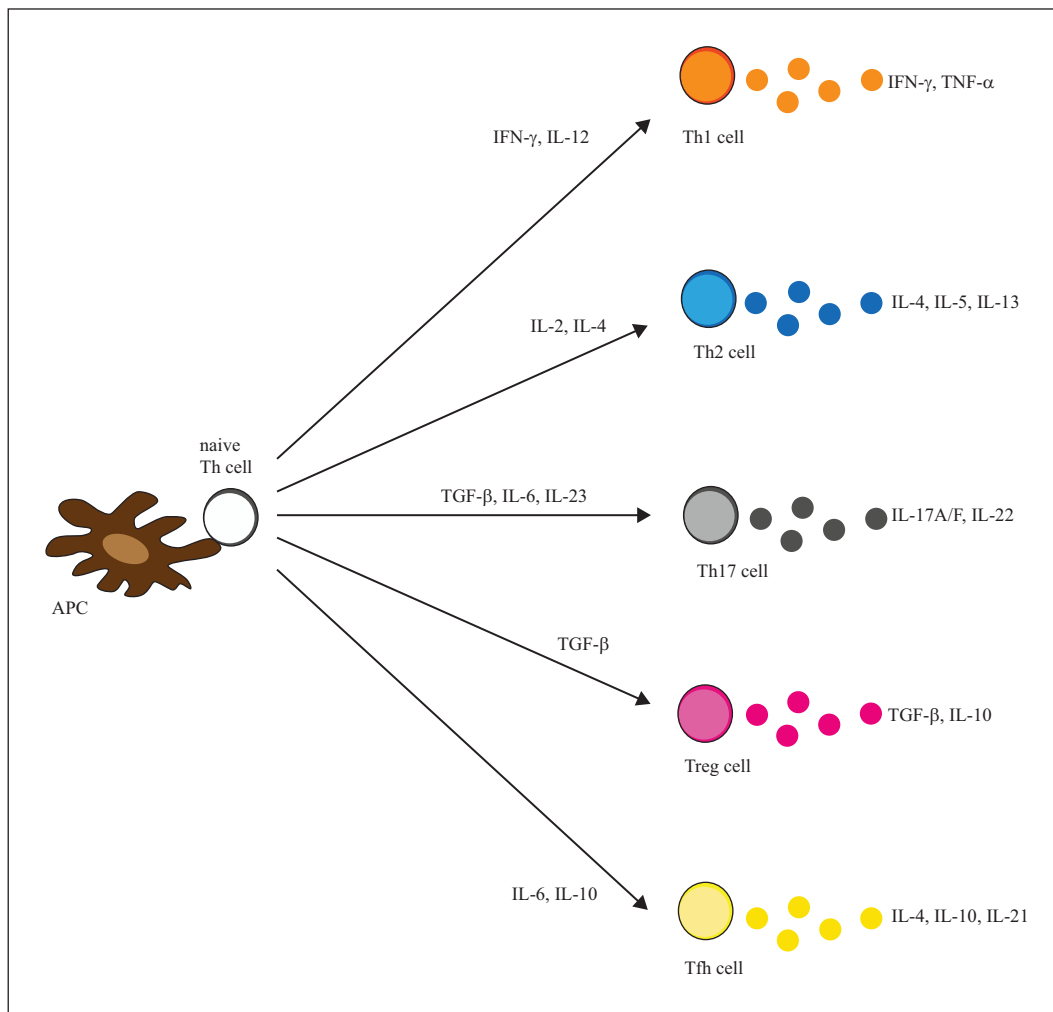


Figure 2

T-helper cell differentiation and key cytokines involved.

APC: antigen-presenting cell; IFN: interferon; IL: interleukin; TFH: T-follicular helper; TGF: transforming growth factor; Th: T helper; TNF: tumor necrosis factor; Treg: regulatory T.

CD4⁺ T cells and cytotoxic T cells, playing a critical role in the differentiation of proinflammatory Th1 and Th17 cells and anti-inflammatory Treg cells [26]. Two important mTOR-dependent metabolic pathways that fuel the elevated energy and biosynthetic demands of effector CD4⁺ and CD8⁺ T cells are aerobic glycolysis and glutaminolysis [27]. Treg cells and Th2 cells, which are typically present in lean adipose tissue, have an anti-inflammatory effect on the surrounding tissue [28, 29]. By contrast, CD8⁺ T cells and Th1 cells that migrate to, and are activated in, obese adipose tissue, promote the proliferation of proinflammatory M1 macrophages [29–31]. *Ex vivo*, these cells from adipose tissue exhibit higher proinflammatory cytokine production following restimulation [32]. It has been shown that an overabundance of glucose and lipids in obese tissue can impact T cell activation and differentiation directly *via* the modulation of nutrient sensor activity [29, 33]. Various studies have demonstrated a glucose-dependent increase in the production of proinflammatory cytokines, such as tumor necrosis factor- α , IL-1 β , and IL-6, as well as the expression of insulin or growth factor receptors and their adaptors, including insulin receptor substrate 1, insulin signal peptide (INSP), also known as insulin receptor (IR) and insulin-like growth factor 1 receptor [34, 35]. Higher cytokine release by

naive T cells is caused by increased chromatin decondensation, which requires glycosylation and product-specific receptor (advanced glycation end product receptor [AGER], also known as receptor for advanced glycation end products [RAGE]) signaling and p38 mitogen-activated protein kinase (MAPK) activity [35]. Hyperglycemia can also directly affect epigenetic modifications in cells by modulating the activity of sirtuins, a group of NAD-dependent protein deacetylases, by increasing histone acetyltransferase and acetyl-CoA expressions for use in histone acetylation [36, 37]. Furthermore, glucose concentration also affects activity of various transcription factors, including liver X receptor, forkhead box protein O1, wingless-related integration site (WNT) signaling, and energy-sensing kinase AMPK in different cell types, which are crucial for effector T cell generation, migration, and memory maturation [27]. Diet rich in saturated fatty acids (SFAs) has been associated with visceral adiposity and the development of type 2 diabetes [38]. SFAs can trigger pancreatic islet β -cell activation *via* toll-like receptor 4 (TLR4), leading to the upregulation of CC-chemokine ligand 2, followed by the recruitment of proinflammatory M1 macrophages to the site [39]. Free fatty acids in turn have been demonstrated to impact Th1-, Th17-, and

Treg-cell differentiation [40]. It has been shown that exposure of CD4⁺ T cells to short-chain fatty acids (SCFAs) promotes Treg-cell differentiation, whereas medium-chain fatty acids and long-chain fatty acids (LCFAs) increase Th1- and Th17-cell differentiation *via* p38 MAPKs and Jun NH2-terminal kinase 1 (JNK1) [41]. It has been observed that acetyl coenzyme A carboxylase 1 (ACC1) expression in memory CD4⁺ T cells from obese individuals correlate with the level of circulating Th17 cells [42]. Deletion of ACC1 inhibits Th17-cell differentiation, which can be restored by adding the exogenous fatty acids [42]. A high-fat diet or direct exposure to SCFAs during immune priming suffices to generate proinflammatory CXCR3-expressing CD4⁺ T-effector memory cells *via* activation of phosphatidylinositol 3 kinase and AKT/protein kinase B (PI3K/AKT) signaling [43]. It is unclear, however, whether these cells are Th1 or Th17 cells, given that CXCR3⁺ T cells are capable of producing both interferon gamma (IFN- γ) and IL-17A/E. The precise molecular mechanism responsible for SCFAs modulation of T cell differentiation is not known, although it involves TLR4 signaling [44]. Increased migration of CXCR3⁺ T cells could intensify inflammation in tissues that regulate metabolic homeostasis, such as adipose tissue, muscle, and liver. In case of Th17-cell differentiation, the process depends on the master transcription factor RAR-related orphan receptor gamma (ROR γ t), a nuclear hormone receptor, whose binding to target DNA response elements is modulated by the availability of cholesterol-derived biosynthetic intermediates [27]. Lipid biosynthesis mediated by sterol regulatory element-binding protein is markedly increased in activated CD4⁺ and CD8⁺ T cells [27]. The resulting increase in cholesterol and fatty acid metabolism in obesity generates metabolites that are crucial for modulation of ROR γ t binding to target gene promoters. In this context, it is worth to note that PPAR γ , a nuclear receptor that binds to ω -polyunsaturated fatty acids, is a central regulator of lipid uptake and storage in numerous cell types, including T cells [27]. PPAR γ has been implicated in both the promotion and inhibition of T cell proliferation [45]. Data indicate that agonists of PPAR γ downmodulate IFN- γ production by CD4⁺ and CD8⁺ T cells, and PPAR γ activity controls the generation of visceral adipose tissue (VAT)-resident Treg cells [45]. Importantly, obesity is associated with a reduced level of VAT-resident Treg cells in adipose tissue [46, 47]. This unique Treg population has been recently defined and its main role is to keep the vast depot of adipose tissue quiescent in terms of inflammation to maximally sensitize insulin signaling [48]. VAT-resident Treg cells comprise 40% to 80% of VAT CD4⁺ T cells, and are slightly distinct from lymphoid tissue-associated Treg cells which comprise only 5% to 15% of CD4⁺ T cells [49]. As mentioned, VAT-resident Treg cells are highly dependent upon PPAR γ [distal to T cell receptor (TCR) signaling] and have a unique transcriptome with many lipid metabolism genes upregulated compared to lymphoid tissue-associated Treg cells [49]. It has been shown that overactivation of mTOR, as happens under the conditions of lost fluctuations in diet and leptin

during nutritional overload, reduces the proliferation of Treg cells, inhibits the differentiation of conventional T cells into Treg cells, and promotes the differentiation of the proinflammatory Th1/Th17 cell lineage, thus leading to defective immunoregulation [50, 51]. It has been shown that a strong activation of PI3K-kinase AKT-mTOR pathway restrains the induction of the transcription factor forkhead box protein 3, a master regulator of Treg cells, consequently inhibiting their generation and proliferation [52]. In line with this evidence, elevated and tonic activity of mTOR and the S6 ribosomal kinase maintained in naive CD4⁺ T cells by genetic manipulation, results in lower levels of Treg cells, which in turn leads to the accumulation of proinflammatory T cells, including autoreactive T cells [53]. Treg cells show high sensitivity to oscillatory fluctuations in leptin and circulating nutrients, and rapidly adapt to microenvironmental changes such as low glucose and high lactate, which is typical for inflamed and ischemic tissues [8]. It seems that mTOR signaling needs to be rapid, oscillatory, and of low or intermediate strength to promote the proliferation and homeostasis of Treg *in vivo* and *in vitro*. Altogether, these data indicate that alterations in serum lipid content and abundance in obese individuals influence T cell differentiation, promoting Th17 and Th1 differentiation. It is also worth noting that a nutrient-mediated activation of innate immune cells residing in adipose tissue can further contribute to an inflammatory microenvironment, additionally amplifying T cell activation [54].

LYMPHOCYTE METABOLISM IN AUTOIMMUNE DISEASES

As mentioned, an obesity-associated overabundance of lipids, glucose, and amino acids impacts T cell activation and differentiation directly *via* the modulation of nutrient sensor activity [33]. Similar changes in lymphocyte metabolism are observed in autoimmunity [55] (*figure 3*).

For instance, an increased glucose uptake by T cells with overexpressed glucose transporter 1 (GLUT1) is sufficient to promote IFN- γ and IL-2 production following *in vitro* stimulation and promotes antibody deposition in glomeruli [56]. Moreover, deletion of GLUT1 reduces glycolytic activity in CD4⁺ T cells and effector CD4⁺ T cell generation, protecting against colitis [57]. Reduced GLUT1 mobilization to the plasma membrane due to increased lysosomal degradation of GLUT1-containing vesicles in WASP homolog-deficient T cells also leads to a reduction in T cell proliferation and decreased severity of experimental autoimmune encephalomyelitis (EAE, the mouse model for MS) [58]. Naive CD4⁺ T cells from lupus-prone mice show higher glycolytic rates than those of age-matched control cells [59].

An increased glycolysis rate is a common feature for both effector CD4⁺ T cells (Th1, Th17) and CD8⁺ CTL cells [60, 61]. It has been demonstrated that inhibition of glucose metabolism in CD4⁺ T cells cultured under Th1/Th17 polarizing conditions leads to significantly reduced differentiation into both lineages, whereas

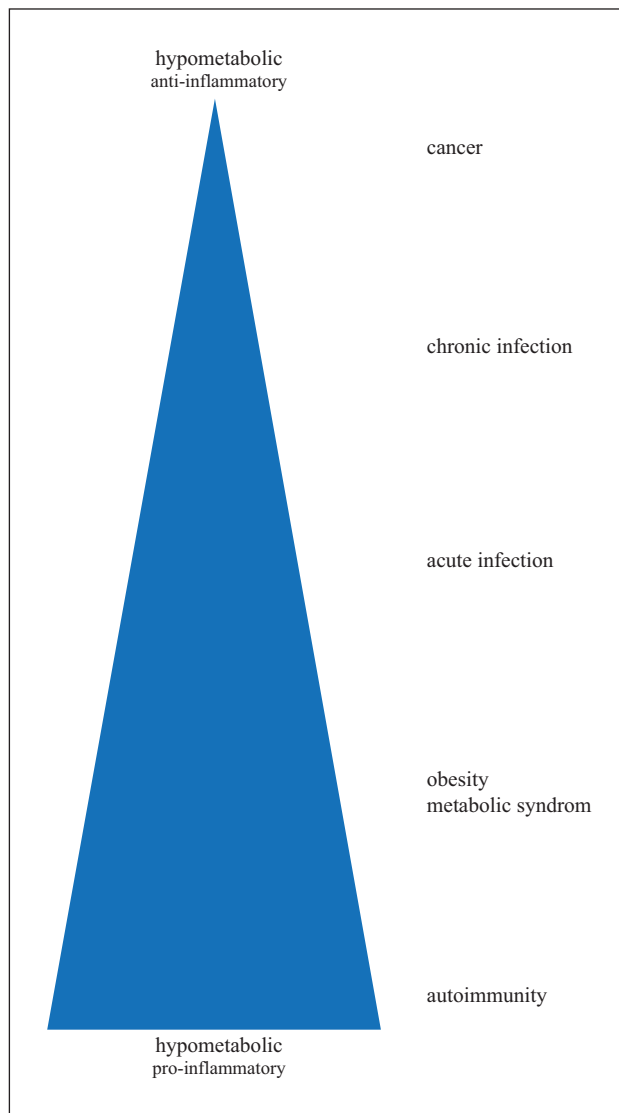


Figure 3
T-cell metabolic states in various diseases.

differentiation into inducible Treg cells is increased under glucose-limiting conditions [60, 62]. Furthermore, blocking glycolysis at the level of hexokinase improves clinical outcome in mice with EAE [60]. It seems therefore that glycolysis is enhanced in all effector T cell populations, but these cells have distinct glucose metabolism signatures.

During an inflammation, immune cells also require increased intracellular pools of amino acids to support broad range of cellular processes, as they are needed as substrates in protein and nucleic acid synthesis, as well as in the synthesis of other amino acids [63, 64]. Increased amino acid availability has been shown to upregulate signal transduction by mTOR1 and to induce stress-response pathways in T cells, whereas restriction differently affects T cell activation, proliferation, and clonal expansion [65]. For instance, halofuginone, an alkaloid that activates the amino acid starvation response by increasing uncharged tRNA levels and activating the amino-acid-sensing kinase general control nonderepressible 2, decreases Th17-cell differentiation [66]. Similar observations have been reported in case of amino acid restriction, including

Met, Cys, Leu, as well as inhibition of tryptophanyl-tRNA charging [66, 67]. Interestingly, neither Th1- nor Th2-cell polarization is affected after activation of the amino acid response pathway, further indicating that effector CD4⁺ Th-cell lineages have distinct sensitivities to amino acid restriction. In line with these findings, halofuginone reduces Th17-cell level in the central nervous system and disease severity in mice with EAE, but does not affect Th1-cell level or IFN- γ production [68]. In addition, leucine and other branched-chain amino acids activate mTORC1 [69]. T cell activation under glutamine-restricted conditions or following inhibition of small neutral amino acid transporters (SNATs) blocks T cell proliferation and IL-2 and IFN- γ production [70, 71]. Glutamine restriction promotes the differentiation of naive CD4⁺ T cells toward Treg-cell lineage, whereas it does not affect the Th2-cell differentiation [72]. In support of these findings, deletion of Slc1a5 which encodes a glutamine transporter, limits Th1- and Th17-cell generation and reduces EAE development [73], whereas SNAT blocking inhibits T cell proliferation and IFN- γ production [71]. Serine deficiency has also been found to significantly reduce proliferation of effector T cells, particularly CD8⁺ T cells [72, 74]. Serine is primarily used in the one-carbon metabolism cycle for purine biosynthesis. It seems plausible that methotrexate (MTX) used for the treatment of various autoimmune diseases, might be responsible for blocking Th17-cell differentiation *via* inhibition of folate-dependent one-carbon metabolism, which is critical for nucleotide biosynthesis [75]. It has been suggested that serine restriction could be used in conjunction with MTX to treat autoimmune diseases such as RA, leading to inhibition of Th17 differentiation and effector function [72].

High levels of circulating lipids associated with obesity might also promote differentiation of CD4⁺ T cells into Th1 and Th17 cells. Rapidly proliferating cells require *de novo* fatty acid and cholesterol synthesis for incorporation into the plasma membrane and lipid membranes of organelles. Targeting glucose-derived *de novo* lipogenesis with an acetyl CoA carboxylase inhibitor blocks Th17 cell differentiation and proliferation, while promoting Treg-cell formation [76]. It is in line with the fact that *de novo* fatty acid and cholesterol synthesis are required for rapidly dividing T cells [72]. It has been found that in a relapsing-remitted model of EAE, linoleic acid and cholesterol metabolism are increased [77]. Dyslipidemia is also observed in individuals with RA, SLE, and MS [55, 78]. It remains unclear how altered lipid abundance promotes autoimmunity. Inhibition of cholesterol synthesis with statins (HMG-CoA reductase inhibitors) has been found to stop EAE progression [79]. Statins can directly modulate immune pathogenesis by inhibiting Th17 cell differentiation while promoting Treg-cell differentiation [80]. Importantly, use of statins was found to lower mortality in patients with RA [81]. It is also worth to note that diet-derived LCFAs and SCFAs have opposite effects on Th17 cell differentiation [41]. LCFAs promote the generation of pathogenic Th1 and Th17 cells, whereas SCFAs promote Treg-cell differentiation. Accordingly, an LCFA-rich diet exacerbates pathology in mice with EAE.

OBESITY AS AN AUTOIMMUNE DISEASE

Obesity itself displays various autoimmune characteristics, in which adipose tissue antigens are targeted by autoreactive T cells [82]. For instance, overweight and obese individuals exhibit: (i) the presence of inflammatory cell infiltrates, including Th1 and Th17 cells, CD8⁺ T cells, and M1-polarized macrophages in adipose tissue which is a target organ; (ii) a reduced level of anti-inflammatory Th2 and Treg cells in the adipose tissue; and (iii) circulating auto-antibodies to adipose tissue and apoptosis of adipocytes [28, 29, 83–85]. The activity of several apokines involved in the control of food intake and Th1 responses, such as leptin, has been associated with autoimmune pathogenesis, including RA, SLE, MS, and T1D [86–88]. As mentioned, obesity often coexists with autoimmune diseases in the same individual, possibly as a result of concurrent analogous mechanisms of preferential differentiation of Th cells into pathogenic Th1 and Th17 cells and altered immunotolerance that operate in that individual but affect different organs.

It has been found that adipose tissue T cells display antigenic bias in their TCR diversity, implying that they undergo prior clonal expansion in response to specific, yet unidentified, autoantigens [29]. It has been suggested that the high levels of oxidized lipids might modify self-proteins and thus generate autoimmune responses [8]. The progression of obesity after ingestion of high-fat diet increases the concentration of “natural” immunoglobulin M (IgM) which results from the activation of TLR4 on B cells [84]. IgM and an apoptosis inhibitor of macrophage (AIM) play an important role in obesity-associated insulin resistance [89]. Higher serum levels of AIM have been found in obese individuals with coexisting autoimmune diseases in sharp contrast to individuals suffering from obesity only, implying that AIM may be used as a potential biomarker for obesity-related autoimmune disorders [84].

As mentioned, evidence suggests that an obesogenic diet significantly contributes to the production of autoantibodies, presumably *via* CD40L signaling, which intensifies the production of inflammatory mediators by adipocytes [90, 91]. Defects in B-cell tolerance have been associated with most of autoimmune diseases and are illustrated by the production of autoantibodies that target self-antigens. Some of these autoantibodies are pathogenic because they interfere with the function of the molecules they recognize, such as the acetylcholine receptor/muscle-specific tyrosine kinase in myasthenia gravis [92], and the aquaporin-4 water channel in neuromyelitis optica spectrum disease [93]. Others target nucleic acids or their associated proteins, allowing the formation of immune complexes that deposit in various organs and induce organ damage, like in systemic SLE [94]. These immune complexes also allow the activation of myeloid cells expressing FcRs-binding autoantibodies and TLRs, including TLR7, TLR8, and TLR9 that recognize autoantibody-bound nucleic acids and lead to cell activation, thereby intensifying inflammation [94]. As B cells have been demonstrated to be essential for the development of diabetes in the NOD mouse model,

additional investigation revealed that B cells promote diabetes by recognizing self-antigens with their autoreactive B-cell receptors (BCRs) and presenting self-antigens *via* MHC class II molecules to T cells [95, 96]. Therefore, self-antigen presentation by autoreactive B cells that escape tolerance may initiate the development of autoimmune diseases [97].

Results from animal and human studies support the concept of pathogenic IgG antibody production directed against specific array of self-antigens in obesity, including: (i) the GOSPR1-Golgi apparatus, which is highly associated with insulin resistance in obese males; (ii) IgG2c antibodies with yet unidentified specificity in mice; and (iii) reduction of B-cell antibody responses in young and elderly individuals with obesity [98, 99]. In addition to the secretion of pathogenic antibodies in VAT [99], a production of autoreactive IgG antibodies in subcutaneous adipose tissue (SAT) in obese individuals has been demonstrated as well. During the development of obesity, reduced oxygen and subsequent hypoxia and cell death all result in the release of proinflammatory cytokines, peptide self-antigens, cell-free DNA, and lipids. Together, these factors may stimulate B-cell class switching and the production of autoimmune IgG antibodies. Fat-specific IgG antibodies are secreted by B cells in the SAT, whereas B cells express mRNA for the transcription factor T-bet and the membrane marker CD11c, both implicated in the production of autoimmune IgG antibodies [99–101].

CONCLUSIONS

The abnormal accumulation of VAT in obesity is associated with metabolic changes that include altered glucose tolerance, insulin resistance, hyperlipidemia, and metabolic syndrome. Obesity also coincides with increased incidence of autoimmune diseases [98], although other factors can influence these changes [102–104]. Accumulating evidence suggest that prolonged metabolic overload related to overnutrition, influenced by genetic and epigenetic components, might affect immunologic self-tolerance through changes in the energy metabolism of immune cells, particularly Treg cells. A strong activation of PI3K-kinase AKT-mTOR axis blocks the induction of transcription factor FOXP3, a master regulator of Treg cells, consequently inhibiting their generation and proliferation, and promoting Th1/Th17 response (metabolic programs regulating other T cell subsets, including Th9 and Th22 are still elusive and require further investigation [26]). Autoimmunity therefore might result from the impaired generation of thymus-derived Treg cells early in life and/or reduced generation of peripheral Treg cells by overstimulation of nutrient-energy sensing pathways. Expanding our knowledge on the topic, particularly on metabolic T-cell flexibility *in vivo*, will provide us with new insights that can be used to develop therapeutic strategies for various inflammatory diseases, including obesity and autoimmune diseases. Targeting specific metabolic pathways is emerging as an important approach to control immune response and maintain the immunologic homeostasis.

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REFERENCES

1. Lee GR. The balance of Th17 *versus* Treg cells in autoimmunity. *Int J Mol Sci* 2018; 19 : 730.
2. MacIver NJ, Michalek RD, Rathmell JC. Metabolic regulation of T lymphocytes. *Annu Rev Immunol* 2013; 31 : 259-83.
3. Cooper GS, Bynum ML, Somers EC. Recent insights in the epidemiology of autoimmune diseases: improved prevalence estimates and understanding of clustering of diseases. *J Autoimmun* 2009; 33 : 197-207.
4. Theofilopoulos AN, Kono DH, Baccala R. The multiple pathways to autoimmunity. *Nat Immunol* 2017; 18 : 716-24.
5. Lerner A, Jeremias P, Matthias T. The world incidence and prevalence of autoimmune diseases is increasing. *Int J Celiac Dis* 2015; 3 : 151-5.
6. Ehlers S, Kaufmann SH, Participants of the 99(th) Dahlem Conference. . Infection, inflammation, and chronic diseases: consequences of a modern lifestyle. *Trends Immunol* 2010; 31 : 184-90.
7. Bach JF. The effect of infections on susceptibility to autoimmune and allergic diseases. *N Engl J Med* 2002; 347 : 911-20.
8. De Rosa V, La Cava A, Matarese G. Metabolic pressure and the breach of immunological self-tolerance. *Nat Immunol* 2017; 18 : 1190-6.
9. Economic Research Service (ERS). *Food availability (per capita) data system*. ERS, 2016. <https://www.ers.usda.gov/data-products/food-availability-per-capita-data-system/>.
10. Manzel A, Muller DN, Hafler DA, Erdman SE, Linker RA, Kleiweietfeld M. Role of “Western diet” in inflammatory autoimmune diseases. *Curr Allergy Asthma Rep* 2014; 14 : 404.
11. Jackson SE, Llewellyn CH, Smith L. The obesity epidemic – Nature *via* nurture: a narrative review of high-income countries. *SAGE Open Med* 2020; 8 : 2050312120918265.
12. Harpsøe MC, Basit S, Andersson M, *et al.* Body mass index and risk of autoimmune diseases: a study within the Danish National Birth Cohort. *Int J Epidemiol* 2014; 43 : 843-55.
13. Gremese E, Tulusso B, Gigante MR, Ferraccioli G. Obesity as a risk and severity factor in rheumatic diseases (autoimmune chronic inflammatory diseases). *Front Immunol* 2014; 5 : 576.
14. Jensen CB, Ängquist LH, Mendall MA, Sørensen TIA, Baker JL, Jess T. Childhood body mass index and risk of inflammatory bowel disease in adulthood: a population-based cohort study. *Am J Gastroenterol* 2018; 113 : 694-701.
15. Odegaard JI, Chawla A. Connecting type 1 and type 2 diabetes through innate immunity. *Cold Spring Harbor Perspect Med* 2012; 2 : a007724.
16. Mokry LE, Ross S, Timpson NJ, Sawcer S, Davey Smith G, Richards JB. Obesity and multiple sclerosis: a Mendelian randomization study. *PLoS Med* 2016; 13 : e1002053.
17. Sterry W, Strober BE, Menter A, International Psoriasis Council. . Obesity in psoriasis: the metabolic, clinical and therapeutic implications. Report of an interdisciplinary conference and review. *Br J Dermatol* 2007; 157 : 649-55.
18. Sikaris KA. The clinical biochemistry of obesity. *Clin Biochem Rev* 2004; 25 : 165-81.
19. Karczewski J, Sledzinska E, Batur A, *et al.* Obesity and inflammation. *Eur Cytokine Netw* 2018; 29 : 83-94.
20. Reilly SM, Saltiel AR. Adapting to obesity with adipose tissue inflammation. *Nat Rev Endocrinol* 2017; 13 : 633-43.
21. Whiteside SK, Snook JP, Williams MA, Weis JJ. Bystander T cells: a balancing act of friends and foes. *Trends Immunol* 2018; 39 : 1021-35.
22. Lercher A, Baazim H, Bergthaler A. Systemic immunometabolism: challenges and opportunities. *Immunity* 2020; 53 : 496-509.
23. Efeyan A, Comb WC, Sabatini DM. Nutrient-sensing mechanisms and pathways. *Nature* 2015; 517 : 301-10.
24. Saxton RA, Sabatini DM. mTOR signaling in growth, metabolism and disease. *Cell* 2017; 168 : 960-76.
25. Chi H. Regulation and function of mTOR signalling in T cell fate decisions. *Nat Rev Immunol* 2012; 12 : 325-38.
26. Kolan SS, Li G, Wik JA, *et al.* Cellular metabolism dictates T cell effector function in health and disease. *Scand J Immunol* 2020; 92 : e12956.
27. Bantug GR, Galluzzi L, Kroemer G, Hess C. The spectrum of T cell metabolism in health and disease. *Nat Rev Immunol* 2018; 18 : 19-34.
28. Winer S, Chan Y, Paltser G, *et al.* Normalization of obesity-associated insulin resistance through immunotherapy. *Nat Med* 2009; 15 : 921-9.
29. Feuerer M, Herrero L, Cipolletta D, *et al.* Lean, but not obese, fat is enriched for a unique population of regulatory T cells that affect metabolic parameters. *Nat Med* 2009; 15 : 930-9.
30. Nishimura S, Manabe I, Nagasaki M, *et al.* CD8+ effector T cells contribute to macrophage recruitment and adipose tissue inflammation in obesity. *Nat Med* 2009; 15 : 914-20.
31. Zúñiga LA, Shen WJ, Joyce-Shaikh B, *et al.* IL-17 regulates adipogenesis, glucose homeostasis, and obesity. *J Immunol* 2010; 185 : 6947-59.
32. Yang H, Youm YH, Vandanmagsar B, *et al.* Obesity increases the production of proinflammatory mediators from adipose tissue T cells and compromises TCR repertoire diversity: implications for systemic inflammation and insulin resistance. *J Immunol* 2010; 185 : 1836-45.
33. Blagih J, Coulombe F, Vincent EE, *et al.* The energy sensor AMPK regulates T cell metabolic adaptation and effector responses *in vivo*. *Immunity* 2015; 42 : 41-54.
34. Stentz FB, Kitabchi AE. Hyperglycemia-induced activation of human T-lymphocytes with *de novo* emergence of insulin receptors and generation of reactive oxygen species. *Biochem Biophys Res Commun* 2005; 335 : 491-5.
35. Martinez N, Vallerskog T, West K, *et al.* Chromatin decondensation and T cell hyperresponsiveness in diabetes-associated hyperglycemia. *J Immunol* 2014; 193 : 4457-68.
36. Chen S, Feng B, George B, Chakrabarti R, Chen M, Chakrabarti S. Transcriptional coactivator p300 regulates glucose-induced gene expression in endothelial cells. *Am J Physiol Endocrinol Metab* 2010; 298 : E127-37.
37. Deb DK, Chen Y, Sun J, Wang Y, Li YC. ATP-citrate lyase is essential for high glucose-induced histone hyperacetylation and fibrogenic gene upregulation in mesangial cells. *Am J Physiol Renal Physiol* 2017; 313 : F423-9.
38. Kennedy A, Martinez K, Chuang CC, LaPoint K, McIntosh M. Saturated fatty acid-mediated inflammation and insulin resistance in adipose tissue: mechanisms of action and implications. *J Nutr* 2009; 139 : 1-4.
39. Eguchi K, Manabe I, Oishi-Tanaka Y, *et al.* Saturated fatty acid and TLR signaling link β cell dysfunction and islet inflammation. *Cell Metab* 2012; 15 : 518-33.

40. Cluxton D, Petrasca A, Moran B, Fletcher JM. Differential regulation of human Treg and Th17 cells by fatty acid synthesis and glycolysis. *Front Immunol* 2019; 10 : 115.
41. Haghighi A, Jörg S, Dusch A, *et al.* Dietary fatty acids directly impact central nervous system autoimmunity *via* the small intestine. *Immunity* 2015; 43 : 817-29.
42. Endo Y, Asou HK, Matsugae N, *et al.* Obesity drives Th17 cell differentiation by inducing the lipid metabolic kinase, ACC1. *Cell Rep* 2015; 12 : 1042-55.
43. Mauro C, Smith J, Cucchi D, *et al.* Obesity-induced metabolic stress leads to biased effector memory CD4⁺ T cell differentiation *via* PI3K p110 δ -Akt-mediated signals. *Cell Metab* 2017; 25 : 593-609.
44. Corrêa-Oliveira R, Fachi JL, Vieira A, Sato FT, Vinolo MAR. Regulation of immune cell function by short-chain fatty acids. *Clin Transl Immunol* 2016; 5 : e73-173.
45. Clark RB, Bishop-Bailey D, Estrada-Hernandez T, Hla T, Puddington L, Padula SJ. The nuclear receptor PPAR gamma and immunoregulation: PPAR gamma mediates inhibition of helper T cell responses. *J Immunol* 2000; 164 : 1364-71.
46. Matarese G, Procaccini C, De Rosa V, Horvath TL, La Cava A. Regulatory T cells in obesity: the leptin connection. *Trends Mol Med* 2010; 16 : 247-56.
47. Cipolletta D, Feuerer M, Li A, *et al.* PPAR- γ is a major driver of the accumulation and phenotype of adipose tissue Treg cells. *Nature* 2012; 486 : 549-53.
48. Wu D, Han JM, Yu X, *et al.* Characterization of regulatory T cells in obese omental adipose tissue in humans. *Eur J Immunol* 2019; 49 : 336-47.
49. Li C, Spallanzani RG, Mathis D. Visceral adipose tissue Tregs and the cells that nurture them. *Immunol Rev* 2020; 295 : 114-25.
50. Delgoffe GM, Pollizzi KN, Waickman AT, *et al.* The kinase mTOR regulates the differentiation of helper T cells through the selective activation of signaling by mTORC1 and mTORC2. *Nat Immunol* 2011; 12 : 295-303.
51. Stelzner K, Herbert D, Popkova Y, *et al.* Free fatty acids sensitize dendritic cells to amplify TH1/TH17-immune responses. *Eur J Immunol* 2016; 46 : 2043-53.
52. Lu L, Barbi J, Pan F. The regulation of immune tolerance by FOXP3. *Nat Rev Immunol* 2017; 17 : 703-17.
53. Daley SR, Coakley KM, Hu DY, *et al.* Rasgrp1 mutation increases naive T-cell CD44 expression and drives mTOR-dependent accumulation of Helios⁺ T cells and autoantibodies. *Elife* 2013; 2 : e01020.
54. Lu J, Zhao J, Meng H, Zhang X. Adipose tissue-resident immune cells in obesity and type 2 diabetes. *Front Immunol* 2019; 10 : 1173.
55. Wu B, Goronzy JJ, Weyand CM. Metabolic fitness of T cells in autoimmune disease. *Immunometabolism* 2020; 2 : e200017.
56. Jacobs SR, Herman CE, Maciver NJ, *et al.* Glucose uptake is limiting in T cell activation and requires CD28-mediated Akt-dependent and independent pathways. *J Immunol* 2008; 180 : 4476-86.
57. Macintyre AN, Gerriets VA, Nichols AG, *et al.* The glucose transporter Glut1 is selectively essential for CD4⁺ T cell activation and effector function. *Cell Metab* 2014; 20 : 61-72.
58. Piotrowski JT, Gomez TS, Schoon RA, Mangalam AK, Billadeau DD. WASH knockout T cells demonstrate defective receptor trafficking, proliferation, and effector function. *Mol Cell Biol* 2013; 33 : 958-73.
59. Yin Y, Choi SC, Xu Z, *et al.* Normalization of CD4⁺ T cell metabolism reverses lupus. *Sci Transl Med* 2015; 7 : 274ra218.
60. Shi LZ, Wang R, Huang G, *et al.* HIF1 α -dependent glycolytic pathway orchestrates a metabolic checkpoint for the differentiation of TH17 and Treg cells. *J Exp Med* 2011; 208 : 1367-76.
61. Wang R, Dillon CP, Shi LZ, *et al.* The transcription factor Myc controls metabolic reprogramming upon T lymphocyte activation. *Immunity* 2011; 35 : 871-82.
62. Michalek RD, Gerriets VA, Jacobs SR, *et al.* Cutting edge: distinct glycolytic and lipid oxidative metabolic programs are essential for effector and regulatory CD4⁺ T cell subsets. *J Immunol* 2011; 186 : 3299-303.
63. Li P, Yin YL, Li D, Kim SW, Wu G. Amino acids and immune function. *Br J Nutr* 2007; 98 : 237-52.
64. Wannemacher RW, Powanda MC, Dinterman RE. Amino acid flux and protein synthesis after exposure of rats to either *Diplococcus pneumoniae* or *Salmonella typhimurium*. *Infect Immun* 1974; 10 : 60-5.
65. Castellano F, Molinier-Frenkel V. Control of T-cell activation and signaling by amino-acid catabolizing enzymes. *Front Cell Dev Biol* 2020; 8 : 613416.
66. Carlson TJ, Pellerin A, Djuretic IM, *et al.* Halofuginone-induced amino acid starvation regulates Stat3-dependent Th17 effector function and reduces established autoimmune inflammation. *J Immunol* 2014; 192 : 2167.
67. Keller TL, Zocco D, Sundrud MS, *et al.* Halofuginone and other febrifugine derivatives inhibit prolyl-tRNA synthetase. *Nat Chem Biol* 2012; 8 : 311-7.
68. Sundrud MS, Koralov SB, Feuerer M, *et al.* Halofuginone inhibits TH17 cell differentiation by activating the amino acid starvation response. *Science* 2009; 324 : 1334-8.
69. Saxton RA, Knockenhauer KE, Wolfson RL, *et al.* Structural basis for leucine sensing by the Sestrin2-mTORC1 pathway. *Science* 2016; 351 : 53-8.
70. Carr EL, Kelman A, Wu GS, *et al.* Glutamine uptake and metabolism are coordinately regulated by ERK/MAPK during T lymphocyte activation. *J Immunol* 2010; 185 : 1037-44.
71. Raposo B, Vaartjes D, Ahlqvist E, Nandakumar KS, Holmdahl R. System A amino acid transporters regulate glutamine uptake and attenuate antibody-mediated arthritis. *Immunology* 2015; 146 : 607-17.
72. Shyer JA, Flavell RA, Bailis W. Metabolic signaling in T cells. *Cell Res* 2020; 30 : 649-59.
73. Nakaya M, Xiao Y, Zhou X, *et al.* Inflammatory T cell responses rely on amino acid transporter ASCT2 facilitation of glutamine uptake and mTORC1 kinase activation. *Immunity* 2014; 40 : 692-705.
74. Ron-Harel N, Santos D, Ghergurovich JM, *et al.* Mitochondrial biogenesis and proteome remodeling promote one-carbon metabolism for T cell activation. *Cell Metab* 2016; 24 : 104-17.
75. Huennekens FM. The methotrexate story: a paradigm for development of cancer chemotherapeutic agents. *Adv Enzyme Regul* 1994; 34 : 397-419.
76. Berod L, Friedrich C, Nandan A, *et al.* *De novo* fatty acid synthesis controls the fate between regulatory T and T helper 17 cells. *Nat Med* 2014; 20 : 1327-33.
77. Mangalam A, Poisson L, Nemutlu E, *et al.* Profile of circulatory metabolites in a relapsing-remitting animal model of multiple sclerosis using global metabolomics. *J Clin Cell Immunol* 2013 Jun 30;4:10.4172/2155-9899.1000150.
78. Bhargava P, Calabresi PA. Metabolomics in multiple sclerosis. *Mult Scler* 2016; 22 : 451-60.

79. Youssef S, Stüve O, Patarroyo JC, *et al.* The HMG-CoA reductase inhibitor, atorvastatin, promotes a Th2 bias and reverses paralysis in central nervous system autoimmune disease. *Nature* 2002; 420 : 78-84.
80. Forero-Peña DA, Gutierrez FRS. Statins as modulators of regulatory T-cell biology. *Mediat Inflamm* 2013; 2013 : 167086.
81. Zeiser R. Immune modulatory effects of statins. *Immunology* 2018; 154 : 69-75.
82. Procaccini C, Carbone F, Galgani M, *et al.* Obesity and susceptibility to autoimmune diseases. *Expert Rev Clin Immunol* 2011; 7 : 287-94.
83. Theofilopoulos AN, Kono DH, Baccala R. The multiple pathways to autoimmunity. *Nat Immunol* 2017; 18 : 716-24.
84. Arai S, Maehara N, Iwamura Y, *et al.* Obesity-associated autoantibody production requires AIM to retain the immunoglobulin M immune complex on follicular dendritic cells. *Cell Rep* 2013; 3 : 1187-98.
85. Winer DA, Winer S, Shen L, *et al.* B cells promote insulin resistance through modulation of T cells and production of pathogenic IgG antibodies. *Nat Med* 2011; 17 : 610-7.
86. Francisco V, Pino J, Campos-Cabaleiro V, *et al.* Obesity, fat mass and immune system: role for leptin. *Front Physiol* 2018; 9 : 640.
87. Procaccini C, Pucino V, Mantzoros CS, Matarese G. Leptin in autoimmune diseases. *Metabolism* 2015; 64 : 92-104.
88. Neumann E, Hasseli R, Ohl S, Lange U, Frommer KW, Müller-Ladner U. Adipokines and autoimmunity in inflammatory arthritis. *Cells* 2021; 10 : 216.
89. Neiman M, Hellström C, Just D, *et al.* Individual and stable autoantibody repertoires in healthy individuals. *Autoimmunity* 2019; 52 : 1-11.
90. Petta I, Fraussen J, Somers V, Kleinewietfeld M. Interrelation of diet, gut microbiome, and autoantibody production. *Front Immunol* 2018; 9 : 439.
91. Tsigalou C, Vallianou N, Dalamaga M. Autoantibody production in obesity: is there evidence for a link between obesity and autoimmunity? *Curr Obes Rep* 2020; 9 : 245-54.
92. Yi JS, Guptill JT, Stathopoulos P, Nowak RJ, O'Connor KC. B cells in the pathophysiology of myasthenia gravis. *Muscle Nerve* 2018; 57 : 172-84.
93. Bennett JL, O'Connor KC, Bar-Or A, *et al.* B lymphocytes in neuromyelitis optica. *Neurol Neuroimmunol Neuroinflamm* 2015; 2 : e104.
94. Tsokos GC, Lo MS, Costa Reis P, Sullivan KE. New insights into the immunopathogenesis of systemic lupus erythematosus. *Nat Rev Rheumatol* 2016; 12 : 716-30.
95. Felton JL, Masada D, Bonami RH, Hulbert C, Thomas JW. Anti-insulin B cells are poised for antigen presentation in type 1 diabetes. *J Immunol* 2018; 201 : 861-73.
96. Hulbert C, Riseili B, Rojas M, Thomas JW. B cell specificity contributes to the outcome of diabetes in nonobese diabetic mice. *J Immunol* 2001; 167 : 5535-8.
97. Meffre E, O'Connor KC. Impaired B-cell tolerance checkpoints promote the development of autoimmune diseases and pathogenic autoantibodies. *Immunol Rev* 2019; 292 : 90-101.
98. Versini M, Jeandel PY, Rosenthal E, Shoenfeld Y. Obesity in autoimmune diseases: not a passive bystander. *Autoimmun Rev* 2014; 13 : 981-1000.
99. Stefan N, Kantartzis K, Machann J, *et al.* Identification and characterization of metabolically benign obesity in humans. *Arch Intern Med* 2008; 168 : 1609-16.
100. Frasca D, Diaz A, Romero M, Vazquez T, Blomberg BB. Obesity induces pro-inflammatory B cells and impairs B cell function in old mice. *Mech Ageing Dev* 2017; 162 : 91-9.
101. Frasca D, Diaz A, Romero M, Thaller S, Blomberg BB. Secretion of autoimmune antibodies in the human subcutaneous adipose tissue. *PLoS One* 2018; 13 : e0197472.
102. Ferrara CT, Geyer SM, Liu YF, *et al.* Excess BMI in childhood: a modifiable risk factor for type 1 diabetes development? *Diabetes Care* 2017; 40 : 698-701.
103. Fourlanos S, Harrison LC, Colman PG. The accelerator hypothesis and increasing incidence of type 1 diabetes. *Curr Opin Endocrinol Diabetes Obes* 2008; 15 : 321-5.
104. Fourlanos S, Varney MD, Tait BD, *et al.* The rising incidence of type 1 diabetes is accounted for by cases with lower-risk human leukocyte antigen genotypes. *Diabetes Care* 2008; 31 : 1546-9.