



# Hypoxic link between cancer cells and the immune system: The role of adenosine and lactate

EDUARDO ALVARADO-ORTIZ<sup>1,2</sup>; MIGUEL ANGEL SARABIA-SÁNCHEZ<sup>3,\*</sup>

<sup>1</sup> Programa de Posgrado en Ciencias Biológicas, Universidad Nacional Autónoma de México, Mexico City, 04510, México

<sup>2</sup> Departamento de Bioquímica, Facultad de Medicina, Universidad Nacional Autónoma de México, Mexico City, 04510, México

<sup>3</sup> Subdirección de Investigación Básica, Instituto Nacional de Cancerología, Secretaría de Salud, Mexico City, 14080, México

**Key words:** Hypoxia, Hypoxia inducible factors (HIF-1/2/3α), Lactate, Adenosine, Immune evasion, Tumor microenvironment (TME)

**Abstract:** The tumor microenvironment (TME) is characterized by a symbiosis between cancer cells and the immune cells. The scarcity of oxygen generates hostility that forces cancer cells to alter their biological features in solid tumors. In response to low oxygen availability, the Hypoxia Inducible Factors (HIF-1/2/3α) act as metabolic mediators, producing extracellular metabolites in the tumor microenvironment that influence the immune cells. The modulation of lactate and adenosine on immune evasion has been widely described; however, under hypoxic conditions, it has been barely addressed. Evidence has demonstrated an interplay between cancer and the immune cells, and the present review explores the findings that support HIFs bridging the gap between the rise of these metabolites and the immunosurveillance failure in a hypoxic context. Moreover, new insights based on systemic oxygen administration are discussed, which might counterbalance the effect mediated by lactate and adenosine, to recover anti-tumor immunity. Thus, the disruption of anti-tumor immunity has been the focus of recent research and this novel avenue opens therapeutic vulnerabilities that can be useful for cancer patients.

## Introduction

The TME (Tumor Microenvironment) is metabolically heterogeneous and commonly displays a differential oxygen gradient [1]. In this context, distinct metabolic sources are used for adaptation, maintenance, and progression under stressful conditions. The rapid proliferation of cancer cells, the less vascularized regions concerning blood vessels, and an abnormal blood vessel structure are causes of hypoxic sites, characterized by being at least 150 μm away from the vasculature, and with a pressure of less than 5 mmHg [2]. The low oxygen availability affects the metabolism of cells located in the hypoxic zone. Indeed, hypoxic TME is related to high extracellular levels of adenosine (up to 100 mM), lactate (up to 40 mM), and acidosis (pH less than 6.8) [3].

In the cancer field, two major avenues of study have emerged depending on the cell type of interest: cancer cells and non-cancerous cells residing in the tumor. The present

review proposes an approach based on the metabolic changes hypoxic cancer cells undergo and how these affect surrounding cells, specifically immune cells, through extracellular metabolites. This approach complements the findings regarding how hypoxia directly triggers immune system evasion, a topic addressed in various previous reviews [4–6].

### *The new insights into the Warburg effect*

One century ago, Otto Warburg discovered that cancer cells maintain a fermentative pathway, in which glucose is metabolized to lactate, even under normoxic conditions. Warburg proposed that cancer cells drive a metabolic rewiring to maintain the metabolic fuels for cancer development. Although the so-called “Warburg effect” represents a less efficient mechanism in terms of ATP generation, the metabolic intermediaries necessary for anabolism are produced [7,8]. Initially, a mitochondrial disruption was the origin of cancer. The current viewpoints on the presence of oncogenes and the loss of tumor suppressor genes as the cause of cancer, with metabolic reprogramming being a consequence, not a cause.

For many years, the Warburg effect was attributed to cancer cells as a unique metabolic state, however, it seems

\*Address correspondence to: Miguel Angel Sarabia-Sánchez, mike\_sarabia@hotmail.com

Received: 26 March 2025; Accepted: 30 May 2025;

Published: 18 July 2025



that this does not necessarily occur. One reason is based on the functionality of mitochondria in subpopulations of cancer cells, due to this organelle being involved in metabolic processes, not limited to ATP production. Furthermore, a metabolic heterogeneity exists within the TME, with some cancer cells having a high glycolytic rate as a source of ATP, while others utilize Oxidative Phosphorylation (OXPHOS) or even a hybrid metabolism, depending on the cellular context [9].

The coexistence of oxidative and glycolytic cells in TME defines the metabolic state of cancer cells, as the waste products of one are the fuel of the other. Thus, multiple cell types within TME cause metabolic compartmentalization, triggering the “Reverse Warburg effect”. In this phenomenon, glycolysis occurs in non-cancerous cells, particularly in Cancer-Associated Fibroblasts (CAFs), which represent one of the most abundant non-malignant cells within solid tumors. The glycolysis-derived lactate is uploaded through Monocarboxylate Transporters (MCT) by the neighboring cancer cells, and employed to produce pyruvate, and subsequently, the synthesis of precursors for anabolism [10]. Glycolysis in CAFs is enhanced because OXPHOS decreases through mitophagy, as a consequence of reactive oxygen species in the TME [11]. Indeed, the release of lactate from CAFs and its incorporation into cancer cells support tumor malignancy [12]. Notably, the intercellular contact involved in the TME communication also led to the reverse Warburg effect [13]. Therefore, different cell types within TME, such as CAFs, transfer metabolites to meet the requirements of cancer cells and promote tumor progression. Because Hypoxia Inducible Factors (HIFs) act to mediate the synthesis, shuttling, and uptake of metabolites, the relationship between HIFs and metabolism is discussed.

#### *Extracellular metabolites and HIFs*

The metabolic plasticity of cancer cells allows them to synthesize metabolic precursors that are necessary to adapt to stressful conditions, such as hypoxia [8,14]. The balance of glycolytic capacities and OXPHOS adjusted upon hypoxic conditions is widely recognized. Therefore, how can the Warburg effect and hypoxia be integrated? It has been argued that hypoxia encourages cancer cells to promote glycolysis, while the Warburg effect (aerobic glycolysis) seems to be favored in regions where oxygen is not a limiting factor [15,16]. Despite this, as mentioned above, metabolic heterogeneity is described in the TME, so how can a metabolic cancer profile be defined that explains the influence on other hallmarks, such as the immune system? One approach to addressing this issue is to consider factors, beyond metabolic enzymes, that emerge under low oxygen conditions, such as HIFs, which provide a complementary outlook. For instance, Glucose Transporter 1 (GLUT-1)-expressing cancer cells were found at the tumor edge, while HIF-1 $\alpha$  was enriched in the tumor center [17].

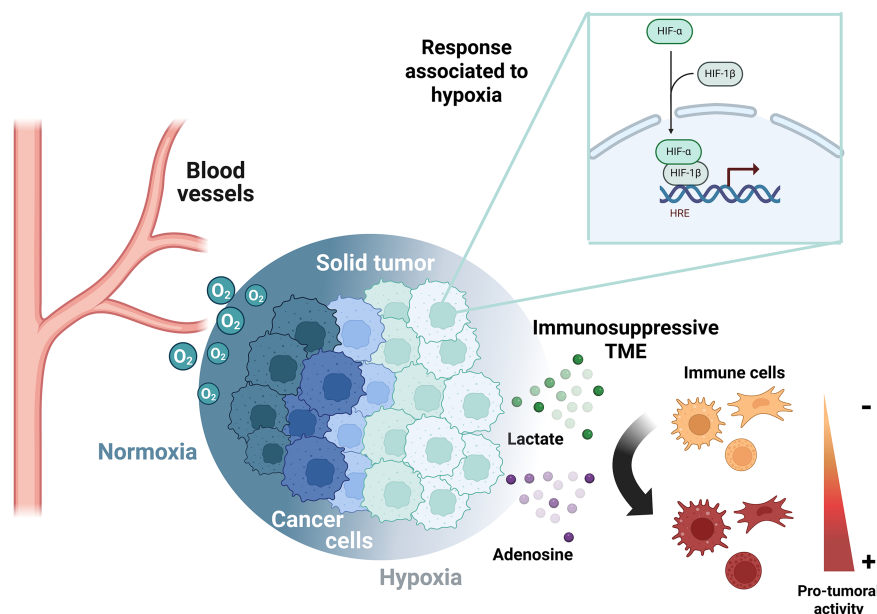
HIF proteins are master transcription factors that regulate multiple responses related to stress conditions, through the formation of heterodimers between HIF- $\alpha$  and HIF- $\beta$  subunits [18]. The function of HIF- $\alpha$  subunits depends on oxygen availability; hence, the transcriptional activity of HIFs is favored in hypoxia. Three HIF- $\alpha$

members have been described: HIF-1 $\alpha$ , HIF-2 $\alpha$  and HIF-3 $\alpha$ . All of these proteins harbor an ODD (Oxygen-Dependent Degradation) domain that is targeted by PHDs (Prolyl Hydroxylase proteins) at proline sites, promoting the degradation of HIFs. The three HIF- $\alpha$  subunits induce gene expression by binding to HIF- $\beta$  (also known as ARNT), which recognizes the Hypoxia Response Element (HRE) consensus sequence in target genes [16,19]. HIF-1 $\alpha$  and HIF-2 $\alpha$  have distinct functions depending on whether hypoxia is acute or chronic. HIF-1 $\alpha$  seems to be active during the acute phase of hypoxia, while HIF-2 $\alpha$  is responsible for adapting to chronic hypoxia [20]. The relevance of HIF-1 $\alpha$  and HIF-2 $\alpha$  has been explored in different types of solid tumors, but the function of HIF-3 $\alpha$  is less well-known. Since HIF-1/2/3 $\alpha$  synchronizes the hypoxic cellular adaptation, none of them should be excluded.

HIF subunits directly regulate the expression profile of metabolic enzymes. HIF-1 $\alpha$  is mainly related to glycolytic response, while HIF-2 $\alpha$  increases transcriptional responses associated with glutamine metabolism dependence or fatty acid synthesis [21–23]. Interestingly, the increased glycolytic rate induced by HIF-1 $\alpha$  is improved under aerobic conditions, suggesting that the functions of HIF-1 $\alpha$  are not limited to a hypoxic microenvironment [24]. Moreover, the mitochondrial activity in cellular adaptation to hypoxia was initially proposed as dispensable, but actually, it is well-known that mitochondria are employed for glutaminolysis and acetate metabolism, suggesting the recovery of additional functions through anaplerotic reactions. In the cases of mitochondria as an energy source, ATP reaches the TME. The extracellular ATP establishes a feedback loop that sustains HIF-1 $\alpha$  activity, enhancing a transcriptional response related to chemoresistance [25]. Strikingly, extracellular ATP in TME originates from various sources, including cell death rates during tumor progression, ongoing exposure to chemotherapeutic drugs, release via specific transporters, and ATP released from extracellular vesicles [26,27]. Metabolic enzymes act on extracellular ATP, leading to the formation of adenosine, which can then act on designated receptors, producing an immunosuppressive state. Notably, different cell types within the TME can express enzymes that promote the accumulation of extracellular adenosine [28].

Target genes of HIF-1 $\alpha$  are related to tumor progression, affecting proliferation, angiogenesis, and metastasis [16]. Also, the tumoral hypoxic zones are described as immune-privileged niches [29]. The extracellular factors modulated by hypoxia and HIF subunits that negatively regulate the immune cells include the overexpression of Vascular Endothelial Growth Factor (VEGF), externalization of phosphatidylserine to the outer membrane, and accumulation of metabolites (adenosine and lactate) within the TME [30]. This is achieved due to the upregulation of the transcriptional activity of HIFs by the low oxygen availability, which indirectly enriches adenosine and lactate in the TME, supporting immune evasion, as will be explained in detail in the review (Fig. 1).

Due to the metabolic reprogramming in cancer, lactate and adenosine coexist in the TME, inducing a plethora of functions in the surrounding cells. Therefore, the metabolic



**FIGURE 1.** Metabolic reprogramming in cancer cells determines immunosuppression in the TME. The oxygen gradient established in solid tumors upregulates HIF- $\alpha$  subunits in hypoxic areas, which restricts the immune system through the release of lactate and adenosine. HRE, Hypoxia response element; TME, tumor microenvironment. The figure was created with BioRender.com

enzymes and transporters associated with these metabolites may be crucial to understanding the effects on immune cells.

### Adenosine in a Hypoxic Microenvironment

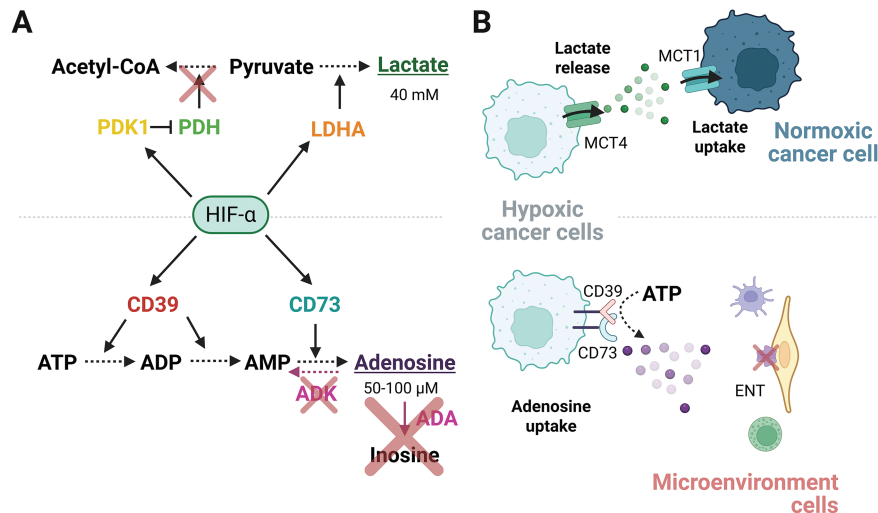
Extracellular ATP accumulates in normal tissues at concentrations ranging from 10 to 100 nM. In tumor tissues, this concentration increases significantly, and extracellular ATP is metabolized by CD39/CD73 ectonucleotidases [31]. Adenosine is synthesized through several enzymatic steps. First, CD39 generates ADP and AMP from ATP. Then, CD73 converts the accumulated AMP into adenosine. Thus, extracellular adenosine in solid tumors primarily originates from extracellular ATP rather than from direct adenosine release [28].

The dynamism of tissue oxygen concentration requires mechanisms that maintain membrane permeability in different physiological processes. Once adenosine is generated, it maintains a basal concentration of around 200 nM. However, if damage occurs, the concentration can reach up to 100  $\mu$ M [32]. Adenosine release occurs at the same time as a reduction in adenosine-metabolizing enzymes [33]. If exposure to chronic hypoxia occurs, it disrupts the blood flow to defined regions, activating ischemia-related processes. Adenosine is well-documented for protecting against damage induced by hypoxia; conversely, the absence of adenosine promotes inflammatory infiltration. Thus, adenosine formation can potentially act as an immune cell regulator within hypoxic sites, modulating vascular permeability [34,35]. Moreover, adenosinergic receptors inhibit pro-inflammatory responses, indicating that adenosine is an important protector against hypoxic damage [36].

Importantly, HIF-1 $\alpha$  regulates the expression of CD39/CD73 ectonucleotidases during hypoxia [37]. Indeed, the promoter region of CD39/CD73 contains an HRE in the

human genome. This was experimentally demonstrated in mice, where liver failure induced HIF-1 $\alpha$  activity and expression of CD39/CD73 [38]. Accordingly, previous reports showed that CD39 and CD73 are regulated by HIF-1 $\alpha$  and identified their binding site in the promoter region [37,39], reinforcing the premise that HIFs are capable of mediating extracellular adenosine accumulation. In this manner, HIF $\alpha$  modulates the adenosine levels, upregulating the expression of CD39 and CD73, which participate in the conversion of ATP to adenosine (Fig. 2A). Although this mechanism is less explored for the other HIFs, the blockade of HIF-2 $\alpha$  disrupts the CD73 expression and the extracellular adenosine accumulation in glioblastoma cells [40]. Furthermore, in triple-negative breast cancer cells, both HIF-1 $\alpha$  and HIF-2 $\alpha$  can bind to the promoter region of CD73 and other genes encoding immunosuppressive molecules, such as PD-L1. The evidence suggests that both HIF $\alpha$  subunits can potentially maintain an immunosuppressive state [41]. Currently, the influence of HIF-2 $\alpha$  and HIF-3 $\alpha$  subunits on the adenosinergic response remains unclear.

Extracellular adenosine is highly unstable, influenced by enzymes that modulate its lifetime through its conversion to other products. For instance, Adenosine Deaminase (ADA) favors the adenosine conversion to inosine, meanwhile, Adenosine kinase (ADK) converts adenosine to AMP through its phosphorylation. The activity of ADA and ADK maintains a compartmentalized concentration of adenosine, coordinating the influence at the extracellular space, or regulating its uptake through specific transporters [42]. The Equilibrative Nucleoside Transporters 1-4 (ENT1-4) belong to a group of proteins involved in mediating the uptake or release of adenosine in a concentration-dependent manner. Hypoxia maintains extracellular adenosine levels by restricting its intracellular uptake or enhancing its release, in a cell-dependent manner. Hypoxia downregulates ENT1,



**FIGURE 2.** Regulation of high levels of lactate and adenosine in the TME. (A) HIF- $\alpha$  subunits transcriptionally upregulate metabolic enzymes that elevate the concentration of lactate and adenosine. (B) Extracellular lactate secreted by hypoxic cancer cells is taken by normoxic cancer cells to use as an energy source, establishing a metabolic symbiosis. Conversely, the downregulation of nucleoside transporters in non-malignant cells prevents the extracellular adenosine uptake released by hypoxic cancer cells. ENT, Equilibrative nucleoside transporters; TME, tumor microenvironment. Figure was created with BioRender.com

which decreases adenosine uptake in umbilical endothelial cells [43]. Therefore, TME cells not only generate adenosine but are also able to capture it, establishing a recycling mechanism to produce energy in the cells that use it. Simultaneously, the responsiveness of non-cancerous cells and non-immune cells decreases, allowing the metabolite to act preferentially on cancer and immune cells (Fig. 2B).

In different types of tissues, hypoxia exposure decreases ADK activity by transcriptional repression mediated by HIF-1 $\alpha$ . This acts as a mechanism to diminish vascular leakage and ischemic tissue damage [44]. However, how is the repressive nature of HIF-1 $\alpha$  explained? This question was explored in the context of cancer, where hypoxia-induced ADK repression maintains the extracellular adenosine concentration necessary to maintain an immunosuppressive microenvironment. Increased activity of the MXI1 repressor, in a HIF1 $\alpha$  mechanism-dependent manner, was involved in this process. Because MXI1 acts upstream of the promoter region of ADK, this mechanism explains why adenosine is not metabolized in liver cancer cells [28]. Together, these results highlight adenosine as an important driver of malignancy, where different enzymatic steps promote its synthesis and prevent its metabolism. Adenosine functions are crucial to tumor malignancy and extend to the TME. Treatments aimed at regulating adenosine concentration have been explored. For example, recombinant ADA reduces the proliferative capacity of cancer cells and inhibits cellular migration induced by hypoxia, through HIF2 $\alpha$  degradation [45].

Furthermore, adenosine receptors are ubiquitously expressed in different cell types, including cancer and non-cancer cells. Thus, the effect of adenosine on the TME is not limited to a single cell type. In this sense, adenosine enhances malignancy through adenosine receptors. For instance, the A2B receptor maintains the “stemness” properties, including tumor initiation capacity and

chemoresistance. Additionally, adenosine promotes proliferation, EMT, invasion, and metastasis in gastric cancer, in an A2A-dependent process [46]. Thus, adenosine receptors are required to mediate malignancy. The functional effects of adenosine receptors on cancer and immune cells have been further explored by other authors [47].

### Lactate in a Hypoxic Microenvironment

In non-malignant cells, the glucose is metabolized to pyruvate through glycolysis. Pyruvate is then oxidized through the TCA cycle to generate the reductive power necessary for OXPHOS. This process produces ~36 molecules of ATP per glucose molecule catabolized [7]. In contrast, cancer cells catalyze the conversion of pyruvate to lactate via the enzyme Lactate Dehydrogenase A (LDHA), regardless of oxygen availability. However, during hypoxia, the conversion of glycolysis-derived pyruvate to lactate rather than to acetyl-CoA is favored. This is achieved because HIF-1 $\alpha$  promotes the expression of LDHA and Pyruvate Dehydrogenase Kinase 1 (PDK1), an enzyme responsible for inhibiting Pyruvate Dehydrogenase (PDH) [48]. The upregulation of PDK1 prevents the oxidation of pyruvate to acetyl-CoA, and it contributes to maintaining the lactate fermentation (Fig. 2A) [49]. In solid tumors, the overall levels of acetyl-CoA are diminished, as is the TCA cycle and OXPHOS, when oxygen is restricted, reducing the supply of ATP through OXPHOS [50]. Notably, the transcription targets regulated by HIF-1 $\alpha$  are consistent with the pro-Warburg effect observed for this subunit, and the main reason for the enrichment of extracellular lactate is the upregulation of LDHA due to improved glycolysis (Warburg effect).

Glutaminolysis also contributes to elevated lactate levels because uptake by the ASCT2 transporter and conversion to  $\alpha$ -ketoglutarate allow entry into the TCA cycle. Then, malate is metabolized to pyruvate by the malic enzyme and



converted to lactate via LDHA. Of note, malic enzymes and LDHA are overexpressed in tumors [51].

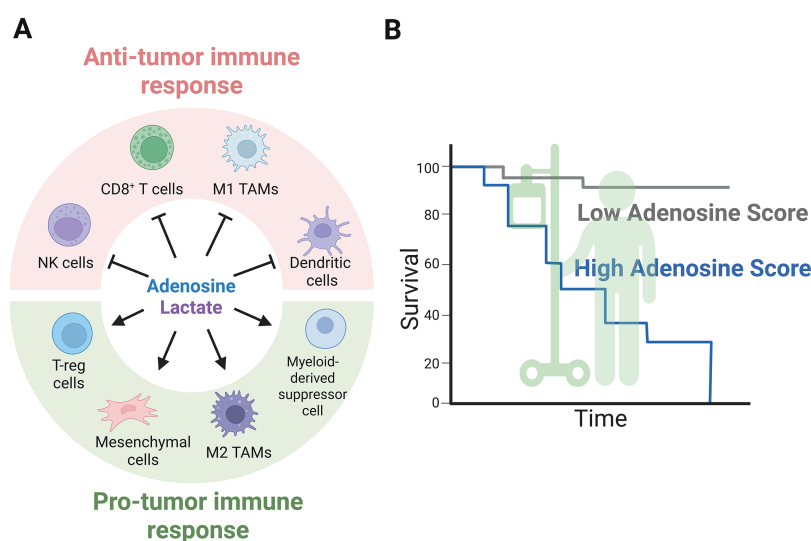
Studies have shown that lactate is heterogeneously distributed in tumors, reaching concentrations of up to 40 mM [52]. The unequal distribution of lactate within the tumor originates from the differential metabolic profiles but is also associated with oxygen availability, as mentioned above. Hypoxic cancer cells favor glucose uptake by upregulating the glucose transporter GLUT-1, thereby increasing glycolysis and, therefore, lactate release. The secreted lactate is then taken up by normoxic cancer cells and converted to pyruvate by LDH-B. In consequence, pyruvate enters the TCA cycle and is used by the mitochondria to produce energy. Hence, lactate is produced in an oxygen-poor zone and utilized in an oxygen-rich zone [53,54]. Additionally, the lactate contributes to the survival of cancer cells located far from the blood vessels [16]. The amount of lactate in the extracellular space depends not only on the metabolic route that produces it but also on the transporters responsible for its mobilization. Specifically, MCT4 mainly drives the lactate release under hypoxic conditions, in contrast to the lactate capture by MCT1 to internalize it into the cells [55,56]. Notably, the export of lactate out of the cell is necessary for oncogenesis, since the inhibition of MCT4 triggers the accumulation of intracellular lactic acid and the impaired survival of the hypoxic cancer cells [57]. Altogether, the hypoxic cancer cells release lactate through MCT4, and subsequently, after diffusion along the TME, lactate is internalized through MCT1 within the normoxic cancer cells. This phenomenon is widely known as metabolic symbiosis, where different cell types obtain a mutual benefit (Fig. 2B).

### The Hypoxia-Adenosine-Immune Evasion Network

The immunosuppressive microenvironment is achieved by adenosine through mechanisms influenced by hypoxia

(Fig. 3A). Identifying adenosine in solid tumors is a challenge that was recently explored. Employing mass spectrometry, researchers found that extracellular adenosine is ubiquitously distributed in pancreatic adenocarcinoma, particularly in hypoxic regions where HIF1 $\alpha$  is present [58]. Adenosine affects the function of different cell types within the TME, including regulatory T-lymphocytes (T-reg), cytotoxic T-lymphocytes, NK cells, and macrophages. Moreover, the CD39/CD73 ectonucleotidases are expressed in immune system cells. Recently, NK cells expressing CD73 with the capacity to reduce the CD4<sup>+</sup> T-lymphocyte population have been observed in breast cancer, suggesting increased extracellular adenosine [59]. However, this population does not synthesize adenosine, suggesting that additional mechanisms may be involved in the immunosuppressive capacities independently of ectonucleotidase activity. In addition, CD73 expression is intrinsically associated with poor clinical outcomes. Thus, adenosine production can act on CD8<sup>+</sup> lymphocytes, causing anergy [60]. Furthermore, mesenchymal cells upregulate CD39 to maintain adenosine synthesis, a process complemented by CD73 expression on T-lymphocytes, while simultaneously decreasing ADA activity [61]. This effect is crucial for reducing the activation of T-lymphocytes, suggesting that the immunosuppressive properties of adenosine are influenced by a network of multiple cell types. Additionally, several reports indicate that hypoxia can affect the paracrine functions of mesenchymal cells, promoting angiogenesis and preventing apoptosis as a protective mechanism in response to damage. However, the role of adenosine in enhancing the malignant properties of cancer cells is not well understood [62,63].

Adenosine generated by a hypoxic microenvironment promotes the enrichment of specific plasmacytoid Dendritic Cells (pDCs). In hepatocellular carcinoma, pDCs are highly infiltrated and play a central role in promoting immunosuppression. Adenosine acts on pDCs, modifying



**FIGURE 3.** Extracellular metabolites mediate anti- and pro-tumor immune responses. (A) Extracellular metabolites such as adenosine and lactate suppress anti-tumor immunity, as illustrated by the actions of NK cells, CD8<sup>+</sup> cells, M1 macrophages, and dendritic cells, while promoting the infiltration of T-reg cells, M2 macrophages, mesenchymal stem cells, and myeloid-derived suppressor cells, all of which possess pro-tumor capabilities. (B) A schematic representation of cancer patient survival linked to high and low adenosinergic signatures explored across various solid tumor types. Figure was created with BioRender.com

immune cells by increasing T-reg cells and decreasing CD8-mediated cytotoxicity [64]. What is the interplay between adenosine, hypoxia, and immune evasion? As described earlier, CD39 expression is directly associated with HIF1 $\alpha$  activity in liver cancer, maintaining the enzymatic conversion of ATP to AMP. This mechanism increases myeloid-derived suppressor cells and prevents their differentiation into dendritic cells. Thus, the immune system suppression is maintained while cytotoxic T-cell responses are downregulated and the T-reg population is augmented, thereby diminishing the anti-tumor immune response. Moreover, the A2A receptor is responsible for this immunosuppressive state; therefore, the use of an adenosine receptor antagonist together with immunotherapy (Anti-PD1) could be a promising clinical approach [28,39,65]. Similar data reports that CD73 expression is intrinsically associated with poor clinical outcomes. In this case, adenosine production acts on CD8<sup>+</sup> lymphocytes, causing energy [60]. Furthermore, HIF1 $\alpha$  induces simultaneous increased expression of CD73, CD47, and PD-L1 in triple-negative breast cancer [41]. This evidence suggests a positive feedback loop in which HIF1 $\alpha$  mediates adenosine production to increase immunosuppression. Additionally, blocking CD73 combined with anti-PD1 immunotherapy promotes CD8<sup>+</sup> tumor infiltration and decreases tumor cell growth in the colorectal cancer model [66]. Therefore, the influence of HIF1 $\alpha$  subunits on the adenosinergic pathway in different cell types of the TME is a promising area of research.

Macrophages are another cell type influenced by adenosine. There exist two types of polarization: anti-tumoral M1 macrophages and protumoral M2 macrophages. M2 macrophages are predominantly found in hypoxic regions, where adenosine is most abundant. The M2 macrophages express the A2A adenosine receptor; therefore, blocking the adenosinergic pathway affects M2 macrophage infiltration into solid tumors, as well as the expression of the PD-L1 immune checkpoint protein. Importantly, the adenosinergic signature in this condition indicates a poor prognosis and survival rate for pancreatic cancer patients (Fig. 3B) [58].

T-reg cells can reduce the exacerbated CD8<sup>+</sup> and CD4<sup>+</sup> lymphocyte function, thereby suppressing anti-tumor immunity in the presence of adenosine and hypoxia [65]. However, the study of both factors is poorly explored. Hypoxic solid tumors show higher T-reg cell infiltration, as previously demonstrated in breast cancer [67]. Similarly, hypoxia induction in a hepatocellular carcinoma model increases T-reg infiltration, which is strongly associated with poor clinical outcomes [68]. In gastric cancer, adenosine synthesized in the TME acts on T-reg cells to suppress the cytolytic capacities of CD8<sup>+</sup> cells and promote their proliferation [69]. Thus, although the individual effects of adenosine and hypoxia on T-reg cells have been explored, both undoubtedly mediate the immunosuppressive characteristics of the TME.

### The Hypoxia-Lactate-Immune Evasion Network

The idea that cancer cells produce lactate for export rather than use it as a nutrient has been discussed [56]. Therefore,

lactate is expected to act as a signal to nearby cells, both cancerous and non-cancerous. The effect of lactate on immune system cells has been a recurrent field of exploration from the perspective of pH regulation [70]. The acidification leads to impairment of anti-tumoral immune cells, for example, modulating the function of NKT and NK, or the recruitment of T-reg cells [71,72]. Lactate release and extracellular acidification are increased in co-cultures of cancer cells and Tumor-Associated Macrophages (TAM), demonstrating the dependence of metabolic profiles on intercellular communication [73]. It should also be taken into account that the pH is modulated by other components, such as proton pumps, which positively regulate the activity of macrophages [74]. Moreover, extracellular acidification was predominantly demonstrated in hypoxia rather than in normoxia, in a glucose-independent manner [75]. Accordingly, glycolysis-deficient cells were able to acidify the extracellular environment [56]. Microenvironment acidification and low oxygen concentration are two important criteria that can influence the immune system cells, and although both are interrelated in the TME, it has been observed that the TAMs are mainly located in hypoxic zones [75].

As mentioned above, lactate is produced in both the absence and presence of oxygen; therefore, it is risky to assume that lactate plays a similar biological role in both conditions. Regulation of lactate on the immune cells has been extensively detailed, but there are few cases where it is considered to occur in a hypoxic context. For instance, a cancer cell-conditioned medium induced M2 macrophage polarization in a normoxic environment, but this effect was amplified when both cancer cells and macrophages were exposed to hypoxia and a higher glucose concentration [75]. Metabolomic screening demonstrated that, while lactate positively correlates with M2 polarization under hypoxic conditions, the metabolic product 2-amino-butanoic acid (2A-BA) correlated with M2 polarization under normoxia [75].

The cytokines and lactate generated by TAMs and cancer cells, respectively, establish a positive feed-forward loop that promotes tumor progression [76]. The pleiotropic functions of lactate and Extracellular Vesicles (EVs) are also implicated in intercellular communication between cancer cells and the immune cells. Among the various components shown to be transported within EVs, long non-coding RNAs (lncRNAs) have recently attracted attention. TAMs release HIF-1 $\alpha$ -stabilizing long noncoding RNA (HISLA), which is contained in EVs. HISLA restricts the binding of PHD2 with HIF-1 $\alpha$ , thereby downregulating the hydroxylation and degradation of HIF-1 $\alpha$ . This is a non-cancer cell-mediated mechanism by which HIF-1 $\alpha$  levels are regulated in cancer cells. Notably, the cancer cell, in turn, produces lactate, which augments HISLA in macrophages, establishing a feed-forward loop. Lactate release is due to metabolic reprogramming caused by HISLA blockade, which impairs glycolysis in cancer cells *in vivo* [73]. Why is this relevant? The Warburg effect refers to glycolysis occurring in the presence of oxygen. Although it may seem counterintuitive that HIFs participate in this phenomenon, evidence from the interaction between the macrophages and cancer cells

suggests a potential role for HIFs in tumor areas beyond hypoxic ones. Notably, the HSLA lncRNA is also expressed in B-lymphocytes, hence, the involvement of additional immune cells in EV-mediated lactate production in cancer cells seems possible.

Importantly, macrophages respond to hypoxic conditions by generating lactate due to their increased glycolytic rate, which mediates histone acetylation and affects the expression of target genes involved in macrophage polarization. This function reveals that lactate has implications for gene expression and favors tumor progression through epigenetic mechanisms [77]. Notably, macrophages have been shown to express HIF-1 $\alpha$ , which requires LDHA activity in cancer cells. This emphasizes the role of lactate and intercellular communication, although it remains to be defined whether lactylation is involved [78]. M2-associated genes are regulated by HIF-1 $\alpha$  [79], so both lactate and HIFs regulate macrophage plasticity, activating or restricting the immune response. However, M2 polarization was associated with the loss of HIF-1 $\alpha$  and overexpression of HIF-2 $\alpha$  [80]. Therefore, the explanation for these contrasting results remains to be elucidated.

TAMs have been found to infiltrate hypoxic areas with a tendency towards immune escape [81]. Additionally, TAMs harboring an M2 phenotype are mainly located in the hypoxic areas of solid tumors [82]. So, then, do HIF subunits mediate macrophage polarization? Interestingly, HIF-1 $\alpha$ -deficient macrophages were recruited to infiltrate hypoxic regions at rates similar to those of macrophages harboring the wild-type version of HIF-1 $\alpha$ . However, HIF-1 $\alpha$ -deficient macrophages exhibited enrichment of M2 phenotype markers and reduced cytotoxicity to tumor cells, suggesting a relationship between HIF subunits and macrophage polarization [80]. Furthermore, HIF-1 $\alpha$  can regulate immune cell infiltration into the tumor. For example, the expression of THBS2, a calcium-associated glycoprotein, was associated with response to immunotherapy. Higher THBS2 expression responded to CTLA4, while lower THBS2 expression showed less immune cell infiltration, but indicated better patient response to anti-PD-L1 therapy. Remarkably, upregulation of THBS2 favored anaerobic metabolism, enhancing lactate secretion. This enrichment of extracellular lactate was reversed when HIF-1 $\alpha$  was downregulated [83]. One of the main mechanisms for lactate action is through GPR132 to modify macrophage polarization [84]. However, infiltration of cytotoxic T-lymphocytes, helper T-lymphocytes, and T-reg cells, which are favored by the overexpression of THBS2, was ablated in mice lacking GPR132. This suggests that THBS2 has a global function that partially depends on the HIF-1 $\alpha$  subunit [83].

It is widely known that cancer cells release a variety of immunosuppressive molecules [85]. In the case of lactate, rather than being a toxic byproduct, it is utilized by the immune cells as part of the adaptive response in hypoxic conditions, but what are the implications of lactate release for immunotherapy? High levels of lactate and hypoxia impair the cytotoxicity mediated by T lymphocytes and modulate T-reg cells [86]. Notably, T-reg cells take up lactate and induce the PD-1 expression to achieve

immunosuppression [86], so the function of lactate in immune checkpoints might be critical for the effectiveness of treatments targeting PD-1.

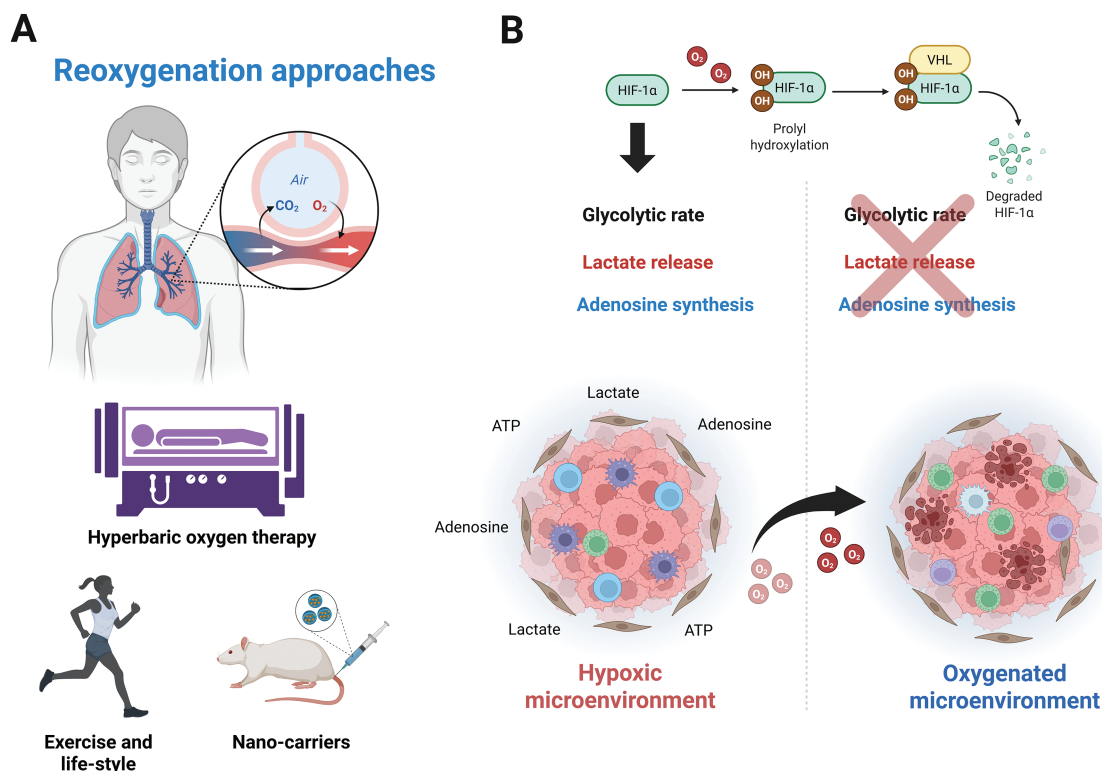
### Adenosine and Lactate: Two Birds in the Same Nest

Adenosine and lactate are enriched in response to hypoxia, however, research aimed at understanding the implications of both metabolites combined is limited. An example of the interconnected role between both metabolites in the same cellular context has been described in cancer cells. Lactate increased the expression of CD38, which is involved in the conversion of NAD<sup>+</sup> to adenosine, through a CD39/CD73-independent mechanism. In this case, the adenosine contributed to invasion and metastasis [87]. Furthermore, the adenosine-producing enzymes CD39/CD73 have been directly associated with the expression of LDH5 and HIF-1 $\alpha$  [88]. In this same report, CD39 in CAFs was related to the expression of PD-L1 and PD-1 on cancer cells and tumor-infiltrating lymphocytes, respectively [88]. These findings recognize that deepening our knowledge of metabolism in the effectiveness of immunotherapy involves considering both cancer cells and the non-cancerous cells that surround them.

In addition to acting as extracellular metabolites, lactate and adenosine regulate epigenetic mechanisms in cancer. The N6-methylation of Adenosine (m<sup>6</sup>A) modulates splicing, stability, and translation of the target RNAs. On the other hand, lactate is involved in histone acetylation to regulate gene transcription. Notably, the lactate-mediated lactylation affects m<sup>6</sup>A modifications to affect the stability of transcripts [89]. Of note, the transcript of lactate transporter MCT1 was described as one of the m<sup>6</sup>A targets, and this regulation enhanced the uptake of lactate to impair cytotoxic CD8<sup>+</sup> lymphocytes [90].

The restoration of the immune cells by immunotherapy leads to immune-related adverse events in some patients. Both adenosine and lactate have been associated with these events in gastrointestinal cancer patients, postulating them as possible prognostic markers for the application of the Immune checkpoint inhibitor [91]. Advances in diagnosis have been made in other diseases. For instance, the measure of LDH and ADA in the pleural fluid has shown an accurate diagnosis of tuberculous pleural effusion [92]. Regarding cancer, the presence of both metabolites in serum has been evaluated as a diagnostic tool in colorectal cancer; however, they exhibited different features. While lactate was useful for contrasting colorectal adenoma with normal tissue, adenosine distinguished between colorectal adenoma and colorectal cancer [93].

As explained, HIFs modulate the amount of lactate and adenosine by acting upstream as a transcriptional regulator, regulating metabolic enzymes. Therefore, by inhibiting the HIF-1 $\alpha$  subunit, the presence of adenosine and lactate is reduced [94]. Due to HIF-1 $\alpha$  blockade abrogating the expression of additional immune system regulators, a function as an immunoadjuvant was considered [94]. Hence, elucidating how adenosine and lactate interplay to control the immune system will undoubtedly provide new therapeutic approaches.



**FIGURE 4.** Therapeutic approaches focused on increasing oxygen infiltration. (A) Oxygen-restoring strategies based on breathing capacity above atmospheric levels explored *in vivo*, such as hyperbaric oxygen therapy, exercise, lifestyle, or nano-carriers. (B) Tumor infiltration and oxygen-dependent mechanisms are downregulated by recovering oxygen in the TME. Figure was created with BioRender.com

### Searching for Therapeutic Options: The Answer Is in the Air

Hypoxia is strongly linked to the disruption of immunosurveillance due to increased adenosine and lactate concentrations. How can this knowledge be useful in therapy? Cells must adapt to a hostile microenvironment to access hypoxic conditions. Recently, hypoxia-modified Chimeric Antigen Receptor (CAR) T-cells have been proposed as a tool to address this issue, by taking advantage of this condition [95]. However, other researchers suggest that oxygen levels can be modulated *in vivo*. Evidence shows that oxygen recovery can restore anti-tumor immunity. To this end, various strategies have been developed, such as nano shuttles or liposomes containing catalase and  $\text{H}_2\text{O}_2$  [96]. Despite substantial evidence supporting the possibility of modulating the oxygen supply, many methods remain methodologically and economically unfeasible. However, the solution may be simpler, as exercise, the lifestyle choices of cancer patients, as well as hyperbaric therapy, could significantly influence the endogenous anti-tumoral response by regulating oxygen concentrations in solid tumors (Fig. 4A).

In mice, hyperoxic breathing at 60%  $\text{O}_2$  decreases the immunosuppression linked to adenosine release, reduces A2A expression, and promotes the recruitment of anti-tumor immune cells, in contrast to breathing at 21%  $\text{O}_2$ . Importantly, while no data is available for cancer patients, in healthy individuals, the reduction in plasma adenosine concentration during exercise is comparable to that achieved when subjects are exposed to a hyperbaric environment (Fig. 4A) [97,98].

Similarly, the ability to target new antigens through machinery associated with Major Histocompatibility Complex (MHC-I) depends on oxygen levels and HIF-1 $\alpha$  activity. Since HIF-1 $\alpha$  acts as a repressor of the MHC-I complex, it has been suggested that when oxygen concentration gradually increases to atmospheric levels (21%  $\text{O}_2$ ) or even exceeds hyperoxic conditions (60%  $\text{O}_2$ ), the capacity for antigen presentation is significantly enhanced [98]. Additionally, hyperoxic exposure reduces components related to the adenosinergic pathway in solid tumors, including extracellular adenosine, CD39/CD73 ectonucleotidases, and adenosine receptors (A2A, A2B). Furthermore, the ability to present antigens through MHC-I was restored, and NK cell-mediated cytotoxicity was improved, thus restricting tumorigenesis and the metastatic capacities of cancer cells [99].

For lactate, although the enrichment of intracellular lactate and the acidification of cancer cells were also observed under 60% oxygen exposure, hyperoxia exposure restrained the tumor growth and metastasis. This phenomenon is attributable to the downregulation of lactate export mediated by MCT1. Hyperoxia and deletion of MCT1 impair glycolysis; however, the function of HIF subunits was not evaluated [100]. Similarly, HIF-1 $\alpha$  is downregulated by hyperbaric oxygen treatment in a manner consistent with degradation mediated by  $\text{O}_2$  availability, which decreases glycolytic flux under low and high glucose conditions. Hyperoxia suppresses the Warburg effect due to the downregulation of key genes involved in glycogen metabolism and glycolysis in a HIF-1 $\alpha$ -dependent manner. Moreover, this report showed that hyperbaric oxygen



therapy reverted the production of pyruvate and lactate by inhibiting the HIF-1 $\alpha$ /Phosphofructokinase Platelet (PFKP) axis [101]. In addition, hyperbaric oxygen therapy reversed LDH activity elicited by hypoxia [102]. Notably, measurements of metabolites in blood samples under hyperoxia-stimulated conditions revealed decreased pyruvate efflux with no effect on PDH activity, highlighting the complexities of this physiological state that require further investigation [103].

The success of therapy depends on the ability to access the hostile environment of hypoxic tumors, which are characterized by high deposition of extracellular matrix. This condition affects the availability of conventional drugs and antibodies used in immunotherapy to act on their targets. For example, in a pancreatic orthotopic mouse model, the combined use of conventional therapies, Abraxane and gemcitabine, along with oxygen recovery, decreases the tumor cell growth by re-establishing the anti-tumoral immune response, reducing the deposition of extracellular matrix, and increasing T-lymphocytes infiltration, as well as the M1/M2 macrophage ratio [104,105]. Similar effects have been observed in other types of solid tumors, in which hyperbaric therapy is effective at eradicating functional properties associated with stemness, enhancing drug uptake within tumors, reducing vascularization, and decreasing the metastatic potential of cancer cells [105]. Despite the benefits of hyperbaric therapy, this approach may be prone to complications. For example, mice treated with doxorubicin are at risk for cardiomyocyte damage. Because systemic exposure to high concentrations of oxygen can also contribute to this dysfunction, as indicated by elevated serum creatine

phosphokinase activity, chemotherapy-induced cardiomyopathy may be slightly exacerbated by hyperbaric oxygen therapy. This has prompted researchers to develop complementary strategies, such as liposome-based approaches, to mitigate the associated side effects [106]. Although hyperoxia does not directly abrogate the tumor cell growth, its effect on the infiltration of antibodies directed to solid tumors without affecting other organs has led to efforts to harness this therapy.

The use of hyperbaric therapy is not without its disadvantages. Recovery of oxygen levels in defined tissues seems to be transient, requiring multiple sessions of hyperoxic breathing to ensure the anti-tumor effect mediated by the immune system [107,108]. Furthermore, various side effects are associated with hyperbaric therapy, including oxygen poisoning, cardiotoxicity, asthma, and barotrauma. Thus, personalized medicine must assess the advantages and disadvantages to determine its usefulness for cancer patients.

Therefore, understanding the influence of oxygen concentration on the secretion of metabolites in cancer cells and its impact on the immune system is crucial for developing new experimental approaches to personalized therapy (Fig. 4B). In this context, a combination of immunotherapy with antagonists of the adenosinergic pathway or lactate production, as well as hyperbaric oxygen therapy could be beneficial for preventing cancer progression and tumor relapse. The use of individual or combined strategies based on the modulation of lactate, adenosine, and oxygen within the TME is currently in clinical trials to overcome hypoxic solid tumors, as listed in Table 1.

TABLE 1

## Clinical strategies targeting regulators of adenosine, lactate and oxygen

| Treatment                                | Target            | Type of cancer  | Observation  | Reference |
|--|-------------------|---|--|-----------|
| Oleclumab                                | CD73              | Non-small cell lung cancer                                | Combined treatment with osimertinib increased the progression-free survival of patients.                               | [109]     |
| Oleclumab                                | CD73              | Colorectal cancer, pancreatic adenocarcinoma, lung cancer | Increased anti-tumor activity and decreased CD73 enzymatic activity.   | [110]     |
| MEDI9447                                 | CD73              | Colorectal cancer   | Increased effector CD8 population and decreased tumor cell growth in mice.   | [111]     |
| CD73-specific siRNA-loaded nanoparticles | CD73              | Mammary carcinoma   | Decreased cell proliferation and sustained tumor regression, lymphangiogenesis in the tumor site and survival of mice. | [112]     |
| Dalutrafusp                              | CD73-TGF- $\beta$ | Solid tumors  | Decreased soluble CD73 and TGF- $\beta$ .  | [113]     |
| AB598                                    | CD39              | Myeloma tumor cells                                       | Increased anti-tumoral immunity and decreased tumor cell growth in mice.   | [114]     |
| TTX030                                   | CD39              | Melanoma cells  | Increased specificity to inhibit CD39 and ADP hydrolysis.  | [115]     |
| MSLN-CAR T-cells secreting anti-CD39     | CD39              | Ovarian cancer cells                                      | Decreased tumor cell growth and increased efficacy of immunotherapy.   | [116]     |

(Continued)

Table 1 (continued)

| Treatment   | Target                                       | Type of cancer                  | Observation   | Reference |
|---|--|---------------------------------|---|-----------|
| 5-aminolevulinic acid (5-ALA)   | LDH  | Glioblastoma                    | Increased cell death in cells dependent on glycolysis.  | [117]     |
| 12-O-deacetyl-phomoxanthone A (12-ODPXA)  | PDK4   | Ovarian cancer                  | Reduced tumor growth and migration <i>in vivo</i> . Suppressed glucose consumption, lactate secretion, and intracellular ATP production.  | [118]     |
| Sononeoperfusion  | Oxygenation                                  | Colon cancer and lung carcinoma | Decreased immunosuppressive TME and increased tumor-infiltrating cytotoxic lymphocytes. Combined treatment with anti-PD-LA enhanced tumor regression and prolonged survival.          | [119]     |
| Cryptotanshinone (KIS37)  | PDK4   | Pancreatic cancer               | Decreased anchorage-independent growth, CSC markers and colony formation. Suppressed cell growth in the orthotopic tumor model.   | [120]     |
| Modified nanoparticles with metabolic inhibitors                                    | Lactate and 6-phosphofructo-2-kinase (PFK-2) | Melanoma                        | Increased immunocompetent TME. Combined treatment with anti-PDL1 decreased tumor growth.  | [121]     |
| PGL13, PGL14, NHI-1 and NHI-2 inhibitors  | GLUT-1 and LDH-A                             | Malignant mesothelioma          | Decreased cell proliferation and cell survival, with a higher effect in combined treatment  | [122]     |
| Metformin   | MCT1 and MCT4                                | Chronic myelogenous leukemia    | Decreased cell proliferation.   | [123]     |
| A2aR-specific siRNA-loaded polyethylene glycol (PEG)-chitosan-lactate nanoparticles | A2aR   | Breast cancer                   | Combined with the Dendritic Cell vaccine, it induced tumor regression and increased survival. Also, combined treatment decreased immunosuppressive cells, angiogenesis and metastasis | [124]     |
| FX11 and AR-C155858   | LDHA and MCT1, respectively                  | Breast and colorectal cancer    | Combined treatment of AR-C155858 and FX-11 decreased proliferation under normoxia or hypoxia  | [125]     |
| Hyperbaric Oxygen Therapy (HBOT)  | O <sub>2</sub>                               | Breast cancer                   | Reduction of tumor cell growth in combination with conventional therapy   | [126]     |
| HBOT  | O <sub>2</sub>                               | Breast cancer                   | Decreased fibrosis associated with cancer disease, without reducing the pain of patients.   | [127]     |
| HBOT + anti-PD1   | O <sub>2</sub>                               | Different types of cell lines   | Enhanced the T-lymphocytes infiltration in TME, decreased the extracellular matrix accumulation, and decreased the tumor cell growth  | [105]     |
| CAR T-cells + Adenosine deaminase   | CD19/HER2                                    | Different types of solid tumors | Enhanced effector activity of CAR T-cells on their target cells in cell lines and mice  | [128,129] |

## Future Perspectives

Enormous progress has been made in describing the metabolic patterns of cancer cells, and it has been demonstrated that non-cancerous cells also undergo metabolic reprogramming through the interplay of extracellular metabolites. This evidence may suggest that research has reached its limits, but it is only the “tip of the iceberg” in a field of knowledge that must be applied to cancer patients. What next? Firstly, research focused predominantly on the HIF-1 $\alpha$  subunit; however, further evidence initiates to recognize that HIF-2 $\alpha$  and HIF-3 $\alpha$  may have an important role in tumor progression. Since the functions of HIF-1 $\alpha$  are not limited to a hypoxic microenvironment, a similar aspect may apply to HIF-2 $\alpha$  and HIF-3 $\alpha$  subunits. Furthermore, the specific use and design of HIF- $\alpha$  inhibitors and their systemic side effects must be carefully addressed.

Research on multiple metabolites is less explored, but a master transcriptional response associated with hypoxia may aid in explaining the extracellular accumulation of both lactate and adenosine. Thus, hypoxia serves as a common driver of their accumulation, making HIF blockade or oxygen recovery a potentially feasible therapeutic strategy in solid tumors. Indeed, a therapeutic strategy that restricts adenosine and lactate is proposed. Understanding the relationship between these metabolites and the immune system, immunotherapy in combination with antagonists of the adenosinergic pathway or lactate production, and hyperbaric oxygen therapy, could be beneficial in preventing cancer progression and tumor relapse. However, clinical trials once again demonstrate that it is necessary to delve deeper into the complexities of a physiological state. Further studies evaluating the synergistic effect of both metabolites on different hallmarks of cancer, including evasion of the

immune system, will allow us to take a key step toward harnessing this knowledge for the benefit of patients.

## Conclusions

Hypoxia is a hallmark of solid tumors characterized by the abundance of extracellular lactate and adenosine. Since oxygen is closely linked to cell metabolism, the effect of HIFs on metabolic reprogramming has predictably been widely discussed [130,131]. A compartmentalized metabolic signature in TME exists, so rather than waste, lactate, and adenosine are master regulators of immune restriction. Advances have addressed additional areas such as the immunological field, suggesting that a balance allows tumor progression [132,133]. The subsequent agreement would undoubtedly require a dialogue between cancerous and non-cancerous cells, within which the metabolites act as messages traveling in both directions.

The scarcity of oxygen, the elevation of adenosine and lactate, as well as the extracellular acidosis, remarkably compromise the immune response [29]. For lactate, the expression of defined MCT1/4 transporters in cancer and immune cells allows for explaining the effect on TME, since for their uptake or release. The alteration of immune cells under hypoxia has been widely addressed [4] and the dual functions of extracellular lactate and adenosine influence the macrophages' polarization, the cytolysis of NK and T-lymphocytes, or the T-reg cell infiltration. Thus, the immune cell infiltration of hypoxic tumors is entirely modified by extracellular metabolites.

The use of metabolic disruptors or adenosinergic antagonists can be beneficial in enhancing the immunotherapy effect. Although *in vitro* and *in vivo* assays sustain this premise, their usefulness in cancer patients needs to be carefully explored. The plasticity of the metabolism in the face of a constantly changing environment explains the difficulty of a comprehensive view, but understanding how metabolites, adenosine, and lactate, in a hypoxic context, alter the immunological state brings us closer to identifying therapeutic hotspots.

**Acknowledgement:** Eduardo Alvarado-Ortiz is a student in the “Doctorado en Ciencias Biológicas” program at the Universidad Nacional Autónoma de México. He received a scholarship from the SECIHTI (Secretaría de Ciencia, Humanidades, Tecnología e Innovación) (CVU854365). Miguel Angel Sarabia-Sánchez received a postdoctoral grant from SECIHTI. The authors express their gratitude to Elizabeth Langley for her constructive comments on the manuscript edition. The figures were created using BioRender.

**Funding Statement:** Not applicable.

**Author Contributions:** Eduardo Alvarado-Ortiz: Writing—review & editing, Writing—original draft, Investigation, Conceptualization. Miguel Angel Sarabia-Sánchez: Writing—review & editing, Writing—original draft, Investigation, Conceptualization. All authors reviewed the results and approved the final version of the manuscript.

**Availability of Data and Materials:** Data sharing is not applicable to this article as no datasets were generated or analyzed during the current study.

**Ethics Approval:** Not applicable.

**Conflicts of Interest:** The authors declare no conflicts of interest to report regarding the present study.

## References

1. Li F, Simon MC. Cancer cells don't live alone: metabolic communication within tumor microenvironments. *Dev Cell*. 2020;54(2):183–95. doi:10.1016/j.devcel.2020.06.018.
2. Al Tameemi W, Dale TP, Al-Jumaily RMK, Forsyth NR. Hypoxia-modified cancer cell metabolism. *Front Cell Dev Biol*. 2019;7:4. doi:10.3389/fcell.2019.00004.
3. Multhoff G, Vaupel P. Radiation-induced changes in microcirculation and interstitial fluid pressure affecting the delivery of macromolecules and nanotherapeutics to tumors. *Front Oncol*. 2012;2:165.
4. Vito A, El-Sayes N, Mossman K. Hypoxia-driven immune escape in the tumor microenvironment. *Cells*. 2020;9(4):992. doi:10.3390/cells9040992.
5. Mortezaee K, Majidpoor J. The impact of hypoxia on immune state in cancer. *Life Sci*. 2021;286(1075–87):120057. doi:10.1016/j.lfs.2021.120057.
6. Peng X, He Y, Huang J, Tao Y, Liu S. Metabolism of dendritic cells in tumor microenvironment: for immunotherapy. *Front Immunol*. 2021;12:613492. doi:10.3389/fimmu.2021.613492.
7. Vander Heiden MG, Cantley LC, Thompson CB. Understanding the Warburg effect: the metabolic requirements of cell proliferation. *Sci*. 2009;324(5930):1029–33. doi:10.1126/science.1160809.
8. Pavlova NN, Zhu J, Thompson CB. The hallmarks of cancer metabolism: still emerging. *Cell Metab*. 2022;34(3):355–77. doi:10.1016/j.cmet.2022.01.007.
9. Fu Y, Liu S, Yin S, Niu W, Xiong W, Tan M, et al. The reverse Warburg effect is likely to be an Achilles' heel of cancer that can be exploited for cancer therapy. *Oncotarget*. 2017;8(34):57813–25. doi:10.18632/oncotarget.18175.
10. Luo M, Luo Y, Mao N, Huang G, Teng C, Wang H, et al. Cancer-associated fibroblasts accelerate malignant progression of non-small cell lung cancer via connexin 43-formed unidirectional gap junctional intercellular communication. *Cell Physiol Biochem Int J Exp Cell Physiol Biochem Pharmacol*. 2018;51(1):315–36. doi:10.1159/000495232.
11. Martinez-Outschoorn UE, Lin Z, Trimmer C, Flomenberg N, Wang C, Pavlides S, et al. Cancer cells metabolically «fertilize» the tumor microenvironment with hydrogen peroxide, driving the Warburg effect: implications for PET imaging of human tumors. *Cell Cycle Georget Tex*. 2011;10(15):2504–20. doi:10.4161/cc.10.15.16585.
12. Sun K, Tang S, Hou Y, Xi L, Chen Y, Yin J, et al. Oxidized ATM-mediated glycolysis enhancement in breast cancer-associated fibroblasts contributes to tumor invasion through lactate as metabolic coupling. *eBioMedicine*. 2019;41:370–83. doi:10.1016/j.ebiom.2024.105542.
13. Fiaschi T, Marini A, Giannoni E, Taddei ML, Gandellini P, De Donatis A, et al. Reciprocal metabolic reprogramming through lactate shuttle coordinately influences tumor-stroma interplay.

- Cancer Res. 2012;72(19):5130–40. doi:10.1158/0008-5472.can-12-1949.
14. DeBerardinis RJ, Lum JJ, Hatzivassiliou G, Thompson CB. The biology of cancer: metabolic reprogramming fuels cell growth and proliferation. *Cell Metab.* 2008;7(1):11–20. doi:10.1016/j.cmet.2007.10.002.
  15. Ibrahim-Hashim A, Robertson-Tessi M, Enriquez-Navas PM, Damaghi M, Balagurunathan Y, Wojtkowiak JW, et al. Defining cancer subpopulations by adaptive strategies rather than molecular properties provides novel insights into intratumoral evolution. *Cancer Res.* 2017;77(9):2242–54.
  16. Missiaen R, Lesner NP, Simon MC. HIF: a master regulator of nutrient availability and metabolic cross-talk in the tumor microenvironment. *EMBO J.* 2023;42(6):1431. doi:10.1525/emboj.2022112067.
  17. Lloyd MC, Cunningham JJ, Bui MM, Gillies RJ, Brown JS, Gatenby RA. Darwinian dynamics of intratumoral heterogeneity: not solely random mutations but also variable environmental selection forces. *Cancer Res.* 2016;76(11):3136–44.
  18. Semenza GL. Pharmacologic targeting of hypoxia-inducible factors. *Annu Rev Pharmacol Toxicol.* 2019;59(1):379–403. doi:10.1146/annurev-pharmtox-010818-021637.
  19. Lee KE, Simon MC. SnapShot: hypoxia-inducible factors. *Cell.* 2015;163(5):1288–1288.e1. doi:10.1016/j.cell.2015.11.011.
  20. Holmquist-Mengelbier L, Fredlund E, Löfstedt T, Noguera R, Navarro S, Nilsson H, et al. Recruitment of HIF-1 $\alpha$  and HIF-2 $\alpha$  to common target genes is differentially regulated in neuroblastoma: HIF-2 $\alpha$  promotes an aggressive phenotype. *Cancer Cell.* 2006;10(5):413–23. doi:10.1016/j.ccr.2006.08.026.
  21. Du W, Zhang L, Brett-Morris A, Aguila B, Kerner J, Hoppel CL, et al. HIF drives lipid deposition and cancer in ccRCC via repression of fatty acid metabolism. *Nat Commun.* 2017;8(1):204. doi:10.1038/s41467-017-01965-8.
  22. Xiao H, Qu Y, Li H, Zhang Y, Fei M, Liang C, et al. HIF-2 $\alpha$ /LINC02609/APOL1-mediated lipid storage promotes endoplasmic reticulum homeostasis and regulates tumor progression in clear-cell renal cell carcinoma. *J Exp Clin Cancer Res CR.* 2024;43(1):209. doi:10.1186/s13046-023-02940-6.
  23. Pérez-Escuredo J, Dadhich RK, Dhup S, Cacace A, Van Hée VF, De Saedeleer CJ, et al. Lactate promotes glutamine uptake and metabolism in oxidative cancer cells. *Cell Cycle Georget Tex.* 2016;15(1):72–83. doi:10.1080/15384101.2015.1120930.
  24. Lu H, Forbes RA, Verma A. Hypoxia-inducible factor 1 activation by aerobic glycolysis implicates the Warburg effect in carcinogenesis. *J Biol Chem.* 2002;277(26):23111–5. doi:10.1074/jbc.m202487200.
  25. Yang H, Geng YH, Wang P, Zhang HQ, Fang WG, Tian XX. Extracellular ATP promotes breast cancer chemoresistance via HIF-1 $\alpha$  signaling. *Cell Death Dis.* 2022;13(3):1–14. doi:10.1038/s41419-022-04647-6.
  26. del Campos-Contreras AR, Díaz-Muñoz M, Vázquez-Cuevas FG. Purinergic signaling in the hallmarks of cancer. *Cells.* 2020;9(7):1612. doi:10.3390/cells9071612.
  27. Shukla S, Dalai P, Agrawal-Rajput R. Metabolic crosstalk: extracellular ATP and the tumor microenvironment in cancer progression and therapy. *Cell Sig.* 2024;121(1):111281. doi:10.1016/j.cellsig.2024.111281.
  28. Cheu JWS, Chiu DKC, Kwan KKL, Yang C, Yuen VWH, Goh CC, et al. Hypoxia-inducible factor orchestrates adenosine metabolism to promote liver cancer development. *Sci Adv.* 2023;9(18):224. doi:10.1126/sciadv.ade5111.
  29. Multhoff G, Vaupel P. Hypoxia compromises anti-cancer immune responses. *Adv Exp Med Biol.* 2020;1232(Suppl 3):131–43. doi:10.1007/978-3-030-34461-0\_18.
  30. Vaupel P, Multhoff G. Hypoxia-/HIF-1 $\alpha$ -driven factors of the tumor microenvironment impeding antitumor immune responses and promoting malignant progression. *Adv Exp Med Biol.* 2018;1072(Suppl 3):171–5. doi:10.1007/978-3-319-91287-5\_27.
  31. Pellegatti P, Raffaghello L, Bianchi G, Piccardi F, Pistoia V, Virgilio FD. Increased level of extracellular ATP at tumor sites: *In Vivo* imaging with plasma membrane luciferase. *PLoS One.* 2008;3(7):e2599. doi:10.1371/journal.pone.0002599.
  32. Layland J, Carrick D, Lee M, Oldroyd K, Berry C. Adenosine: physiology, pharmacology, and clinical applications. *JACC Cardiovasc Interv.* 2014;7(6):581–91.
  33. Kobayashi S, Zimmermann H, Millhorn DE. Chronic hypoxia enhances adenosine release in rat PC12 cells by altering adenosine metabolism and membrane transport. *J Neurochem.* 2000;74(2):621–32. doi:10.1046/j.1471-4159.2000.740621.x.
  34. Eltzschig HK, Ibla JC, Furuta GT, Leonard MO, Jacobson KA, Enjyoji K, et al. Coordinated adenosine nucleotide phosphohydrolysis and nucleoside signaling in posthypoxic endothelium. *J Exp Med.* 2003;198(5):783–96. doi:10.1084/jem.20030891.
  35. Thompson LF, Eltzschig HK, Ibla JC, Van De Wiele CJ, Resta R, Morote-Garcia JC, et al. Crucial role for ecto-5'-nucleotidase (CD73) in vascular leakage during hypoxia. *J Exp Med.* 2004;200(11):1395–405. doi:10.1084/jem.20040915.
  36. Bowser JL, Lee JW, Yuan X, Eltzschig HK. The hypoxia-adenosine link during inflammation. *J Appl Physiol.* 2017;123(5):1303–20.
  37. Synnestvedt K, Furuta GT, Comerford KM, Louis N, Karhausen J, Eltzschig HK, et al. Ecto-5'-nucleotidase (CD73) regulation by hypoxia-inducible factor-1 mediates permeability changes in intestinal epithelia. *J Clin Invest.* 2002;110(7):993–1002. doi:10.1172/jci0215337.
  38. Tak E, Jung DH, Kim SH, Park GC, Jun DY, Lee J, et al. Protective role of hypoxia-inducible factor-1 $\alpha$ -dependent CD39 and CD73 in fulminant acute liver failure. *Toxicol Appl Pharmacol.* 2017;314:72–81. doi:10.1016/j.taap.2016.11.016.
  39. Chiu DKC, Tse APW, Xu IMJ, Di Cui J, Lai RKH, Li LL, et al. Hypoxia inducible factor HIF-1 promotes myeloid-derived suppressor cells accumulation through ENTPD2/CD39L1 in hepatocellular carcinoma. *Nat Commun.* 2017;8(1):234. doi:10.1038/s41467-017-00530-7.
  40. Torres Á, Erices JI, Sanchez F, Ehrenfeld P, Turchi L, Virolle T, et al. Extracellular adenosine promotes cell migration/invasion of glioblastoma stem-like cells through A3 adenosine receptor activation under hypoxia. *Cancer Lett.* 2019;446(6):112–22. doi:10.1016/j.canlet.2019.01.004.
  41. Samanta D, Park Y, Ni X, Li H, Zahnow CA, Gabrielson E, et al. Chemotherapy induces enrichment of CD47+/CD73+/PDL1+ immune evasive triple-negative breast cancer cells. *Proc Natl Acad Sci U S A.* 2018;115(6):E1239–48. doi:10.1073/pnas.1718197115.
  42. Losenkova K, Zuccarini M, Karikoski M, Laurila J, Boison D, Jalkanen S, et al. Compartmentalization of adenosine metabolism in cancer cells and its modulation during acute hypoxia. *J Cell Sci.* 2020;133(10):jcs241463. doi:10.1242/jcs.241463.



43. Casanello P, Torres A, Sanhueza F, González M, Fariás M, Gallardo V, et al. Equilibrative nucleoside transporter 1 expression is downregulated by hypoxia in human umbilical vein endothelium. *Circ Res.* 2005;97(1):16–24. doi:10.1161/01.res.0000172568.49367.f8.
44. Morote-García JC, Rosenberger P, Kuhlicke J, Eltzschig HK. HIF-1-dependent repression of adenosine kinase attenuates hypoxia-induced vascular leak. *Blood.* 2008;111(12):5571–80. doi:10.1182/blood-2007-11-126763.
45. Niechi I, Uribe-Ojeda A, Erices JI, Torres Á, Uribe D, Rocha JD, et al. Adenosine depletion as a new strategy to decrease glioblastoma stem-like cells aggressiveness. *Cells.* 2019;8(11):1353. doi:10.3390/cells8111353.
46. Shi L, Wu Z, Miao J, Du S, Ai S, Xu E, et al. Adenosine interaction with adenosine receptor A2a promotes gastric cancer metastasis by enhancing PI3K-AKT-mTOR signaling. 2019;30(19):2527–34.
47. Fishman P, Bar-Yehuda S, Synowitz M, Powell JD, Klotz KN, Gessi S, et al. Adenosine receptors and cancer. *Handb Exp Pharmacol.* 2009;193(Suppl 1):399–441. doi:10.1007/978-3-540-89615-9\_14.
48. Whan KJ, Tchernyshyov I, Semenza GL, Dang CV. HIF-1-mediated expression of pyruvate dehydrogenase kinase: a metabolic switch required for cellular adaptation to hypoxia. *Cell Metab.* 2006;3(3):177–85. doi:10.1016/j.cmet.2006.02.002.
49. Taylor CT, Scholz CC. The effect of HIF on metabolism and immunity. *Nat Rev Nephrol.* 2022;18(9):573–87. doi:10.1038/s41581-022-00587-8.
50. Papandreou I, Lim AL, Laderoute K, Denko NC. Hypoxia signals autophagy in tumor cells via AMPK activity, independent of HIF-1, BNIP3, and BNIP3L. *Cell Death Differ.* 2008;15(10):1572–81. doi:10.1038/cdd.2008.84.
51. Nakashima C, Kirita T, Yamamoto K, Mori S, Luo Y, Sasaki T, et al. Malic enzyme 1 is associated with tumor budding in oral squamous cell carcinomas. *Int J Mol Sci.* 2020;21(19):7149. doi:10.3390/ijms21197149.
52. Vaupel P. Tumor microenvironmental physiology and its implications for radiation oncology. *Semin Radiat Oncol.* 2004;14(3):198–206. doi:10.1016/j.semradonc.2004.04.008.
53. Feron O. Pyruvate into lactate and back: from the Warburg effect to symbiotic energy fuel exchange in cancer cells. *Radiother Oncol J Eur Soc Ther Radiol Oncol.* 2009;92(3):329–33. doi:10.1016/j.radonc.2009.06.025.
54. Sonveaux P, Végran F, Schroeder T, Wergin MC, Verrax J, Rabbani ZN, et al. Targeting lactate-fueled respiration selectively kills hypoxic tumor cells in mice. *J Clin Invest.* 2008;118(12):3930–42. doi:10.1172/jci36843.
55. Draoui N, Feron O. Lactate shuttles at a glance: from physiological paradigms to anti-cancer treatments. *Dis Model Mech.* 2011;4(6):727–32. doi:10.1242/dmm.007724.
56. de la Cruz-López KG, Castro-Muñoz LJ, Reyes-Hernández DO, García-Carrancá A, Manzo-Merino J. Lactate in the regulation of tumor microenvironment and therapeutic approaches. *Front Oncol.* 2019;9:1143. doi:10.3389/fonc.2019.01143.
57. Morais-Santos F, Granja S, Miranda-Gonçalves V, Moreira AHJ, Queirós S, Vilaça JL, et al. Targeting lactate transport suppresses in vivo breast tumour growth. *Oncotarget.* 2015;6(22):19177–89. doi:10.18632/oncotarget.3910.
58. Graziano V, Dannhorn A, Hulme H, Williamson K, Buckley H, Karim SA, et al. Defining the spatial distribution of extracellular adenosine revealed a myeloid-dependent immunosuppressive microenvironment in pancreatic ductal adenocarcinoma. *J Immunother Cancer.* 2023;11(8):e006457. doi:10.1101/2022.05.24.493238.
59. Neo SY, Yang Y, Record J, Ma R, Chen X, Chen Z, et al. CD73 immune checkpoint defines regulatory NK cells within the tumor microenvironment. *J Clin Invest.* 2020;130(3):1185–98. doi:10.1172/jci128895.
60. Loi S, Pommey S, Haibe-Kains B, Beavis PA, Darcy PK, Smyth MJ, et al. CD73 promotes anthracycline resistance and poor prognosis in triple negative breast cancer. *Proc Natl Acad Sci.* 2013;110(27):11091–6. doi:10.1073/pnas.122251110.
61. Saldanha-Araujo F, Ferreira FIS, Palma PV, Araujo AG, Queiroz RHC, Covas DT, et al. Mesenchymal stromal cells up-regulate CD39 and increase adenosine production to suppress activated T-lymphocytes. *Stem Cell Res.* 2011;7(1):66–74. doi:10.1016/j.scr.2011.04.001.
62. Xia X, Chiu PWY, Lam PK, Chin WC, Ng EKW, Lau JYW. Secretome from hypoxia-conditioned adipose-derived mesenchymal stem cells promotes the healing of gastric mucosal injury in a rodent model. *Biochim Biophys Acta Mol Basis Dis.* 2018;1864(1):178–88. doi:10.1016/j.bbadis.2017.10.009.
63. Yang Y, Lee EH, Yang Z. Hypoxia-conditioned mesenchymal stem cells in tissue regeneration application. *Tissue Eng Part B Rev.* 2022;28(5):966–77. doi:10.1089/ten.teb.2021.0145.
64. Pang L, Ng KTP, Liu J, Yeung WHO, Zhu J, Chiu TLS, et al. Plasmacytoid dendritic cells recruited by HIF-1 $\alpha$ /eADO/ADORA1 signaling induce immunosuppression in hepatocellular carcinoma. *Cancer Lett.* 2021;522(2):80–92. doi:10.1016/j.canlet.2021.09.022.
65. Ohue Y, Nishikawa H. Regulatory T (Treg) cells in cancer: can Treg cells be a new therapeutic target? *Cancer Sci.* 2019;110(7):2080–9. doi:10.1111/cas.14069.
66. Lian W, Jiang D, Lin W, Jiang M, Zhang Y, Wang H, et al. Dual role of CD73 as a signaling molecule and adenosine-generating enzyme in colorectal cancer progression and immune evasion. *Int J Biol Sci.* 2024;20(1):137–51. doi:10.7150/ijbs.87440.
67. Ren L, Yu Y, Wang L, Zhu Z, Lu R, Yao Z. Hypoxia-induced CCL28 promotes recruitment of regulatory T cells and tumor growth in liver cancer. *Oncotarget.* 2016;7(46):75763–73. doi:10.18632/oncotarget.12409.
68. Yan M, Jene N, Byrne D, Millar EKA, O'Toole SA, McNeil CM, et al. Recruitment of regulatory T cells is correlated with hypoxia-induced CXCR4 expression, and is associated with poor prognosis in basal-like breast cancers. *Breast Cancer Res BCR.* 2011;13(2):R47.
69. Shi L, Feng M, Du S, Wei X, Song H, Yixin X, et al. Adenosine generated by regulatory T cells induces CD8+ T cell exhaustion in gastric cancer through A2aR pathway. *BioMed Res Int.* 2019;2019(10142):4093214. doi:10.1155/2019/4093214.
70. Singh L, Nair L, Kumar D, Arora MK, Bajaj S, Gadewar M, et al. Hypoxia induced lactate acidosis modulates tumor microenvironment and lipid reprogramming to sustain the cancer cell survival. *Front Oncol.* 2023;13:1034205. doi:10.3389/fonc.2023.1034205.
71. Pollizzi KN, Powell JD. Integrating canonical and metabolic signalling programmes in the regulation of T cell responses. *Nat Rev Immunol.* 2014;14(7):435–46. doi:10.1038/nri3701.
72. Li W, Tanikawa T, Kryczek I, Xia H, Li G, Wu K, et al. Aerobic glycolysis controls myeloid-derived suppressor cells and tumor immunity via a specific CEBPB isoform in triple-negative breast cancer. *Cell Metab.* 2018;28(1):87–103.e6. doi:10.1016/j.cmet.2018.04.022.

73. Chen F, Chen J, Yang L, Liu J, Zhang X, Zhang Y, et al. Extracellular vesicle-packaged HIF-1 $\alpha$ -stabilizing lncRNA from tumour-associated macrophages regulates aerobic glycolysis of breast cancer cells. *Nat Cell Biol.* 2019; 21(4):498–510. doi:10.1038/s41556-019-0299-0.
74. Huber V, Camisaschi C, Berzi A, Ferro S, Lugini L, Triulzi T, et al. Cancer acidity: an ultimate frontier of tumor immune escape and a novel target of immunomodulation. *Semin Cancer Biol.* 2017;43(4):74–89. doi:10.1016/j.semcancer.2017.03.001.
75. He Y, Xu H, Liu Y, Kempa S, Vechiatto C, Schmidt R, et al. The effects of hypoxia on the immune-metabolic interplay in liver cancer. *Biomolecules.* 2024;14(8):1024. doi:10.3390/biom14081024.
76. Su S, Liu Q, Chen J, Chen J, Chen F, He C, et al. A positive feedback loop between mesenchymal-like cancer cells and macrophages is essential to breast cancer metastasis. *Cancer Cell.* 2014;12(5):605–20. doi:10.1016/j.ccr.2014.03.021.
77. Zhang Y, Peng Q, Zheng J, Yang Y, Zhang X, Ma A, et al. The function and mechanism of lactate and lactylation in tumor metabolism and microenvironment. *Genes Dis.* 2023;10(5):2029–37. doi:10.1016/j.gendis.2022.10.006.
78. Zhang L, Li S. Lactic acid promotes macrophage polarization through MCT-HIF1 $\alpha$  signaling in gastric cancer. *Exp Cell Res.* 2020;388(2):111846.
79. Colegio OR, Chu NQ, Szabo AL, Chu T, Rhebergen AM, Jairam V, et al. Functional polarization of tumour-associated macrophages by tumour-derived lactic acid. *Nat.* 2014;513(7519):559–63. doi:10.1038/nature13490.
80. Werno C, Menrad H, Weigert A, Dehne N, Goerdt S, Schledzewski K, et al. Knockout of HIF-1 $\alpha$  in tumor-associated macrophages enhances M2 polarization and attenuates their pro-angiogenic responses. *Carcinogenesis.* 2010;31(10):1863–72. doi:10.1093/carcin/bgq088.
81. Yang M, McKay D, Pollard JW, Lewis CE. Diverse functions of macrophages in different tumor microenvironments. *Cancer Res.* 2018;78(19):5492–503.
82. Movahedi K, Laoui D, Gysemans C, Baeten M, Stangé G, Van den Bossche J, et al. Different tumor microenvironments contain functionally distinct subsets of macrophages derived from Ly6C(high) monocytes. *Cancer Res.* 2010;70(14):5728–39. doi:10.1158/0008-5472.can-09-4672.
83. Liu Y, Jiang C, Xu C, Gu L. Systematic analysis of integrated bioinformatics to identify upregulated THBS2 expression in colorectal cancer cells inhibiting tumour immunity through the HIF1A/Lactic Acid/GPR132 pathway. *Cancer Cell Int.* 2023;23(1):253. doi:10.1186/s12935-023-03103-5.
84. Chen P, Zuo H, Xiong H, Kolar MJ, Chu Q, Saghatelian A, et al. Gpr132 sensing of lactate mediates tumor-macrophage interplay to promote breast cancer metastasis. *Proc Natl Acad Sci U S A.* 2017;114(3):580–5. doi:10.1073/pnas.1614035114.
85. Zheng Y, Chen H, Zhao Y, Zhang X, Liu J, Pan Y, et al. Knockdown of FBXO22 inhibits melanoma cell migration, invasion and angiogenesis via the HIF-1 $\alpha$ /VEGF pathway. *Invest New Drugs.* 2020;38(1):20–8. doi:10.1007/s10637-019-00761-z.
86. Kumagai S, Koyama S, Itahashi K, Tanegashima T, Lin YT, Togashi Y, et al. Lactic acid promotes PD-1 expression in regulatory T cells in highly glycolytic tumor microenvironments. *Cancer Cell.* 2022;40(2):201–18.e9. doi:10.1016/j.ccell.2022.01.001.
87. Lu Y, Yang Y, Chang T, Jiang Q, Yang C, Fu C, et al. Lactate drives CD38 signaling to promote epithelial-mesenchymal transition through Snail induction in non-small cell lung cancer cells. *J Cell Commun Sig.* 2024;18(1):e12018. doi:10.1002/ccs3.12018.
88. Giatromanolaki A, Kouroupi M, Pouliliou S, Mitrakas A, Hasan F, Pappa A, et al. Ectonucleotidase CD73 and CD39 expression in non-small cell lung cancer relates to hypoxia and immunosuppressive pathways. *Life Sci.* 2020;259:118389. doi:10.1016/j.lfs.2020.118389.
89. Tang F, Xiao D, Li X, Qiao L. The roles of lactate and the interplay with m6A modification in diseases. *Cell Biol Toxicol.* 2024;2(1):107. doi:10.1007/s10565-024-09951-9.
90. Xiong J, He L, Chai X, Zhang Y, Sun S. YTHDF1 boosts the lactate accumulation to potentiate cervical cancer cells immune escape. *Cell Death Dis.* 2024;15(11):127. doi:10.1038/s41419-024-07128-0.
91. Xing N, Liu J, Hou L, Zhao Y, Ma H, Wang F, et al. Clinical biomarkers for thyroid immune-related adverse events in patients with stage III and IV gastrointestinal tumors. *Front Immunol.* 2024;15:1381061. doi:10.3389/fimmu.2024.1381061.
92. Zhao T, Zhang J, Zhang X, Wang C. Clinical significance of pleural fluid lactate dehydrogenase/adenosine deaminase ratio in the diagnosis of tuberculous pleural effusion. *BMC Pulm Med.* 2024;24(1):E486. doi:10.1186/s12890-024-03055-0.
93. Abu Bakar MF, Chin SF, Makpol S, Tan JK, Mohammed Nawi A. Diagnostic performance of serum metabolites biomarker associated with colorectal adenoma: a systematic review. *PeerJ.* 2024;12(11):e18043. doi:10.7717/peerj.18043.
94. Zhang X, He C, He X, Fan S, Ding B, Lu Y, et al. HIF-1 inhibitor-based one-stone-two-birds strategy for enhanced cancer chemodynamic-immunotherapy. *J Control Release Off J Control Release Soc.* 2023;356:649–62. doi:10.1016/j.jconrel.2023.03.026.
95. Kosti P, Opzoomer JW, Larios-Martinez KI, Henley-Smith R, Scudamore CL, Okesola M, et al. Hypoxia-sensing CAR T cells provide safety and efficacy in treating solid tumors. *Cell Rep Med.* 2021;2(4):100227. doi:10.1016/j.xcrm.2021.100227.
96. Feng L, Betzer O, Tao D, Sadan T, Popovtzer R, Liu Z. Oxygen nanoshuttles for tumor oxygenation and enhanced cancer treatment. *CCS Chem.* 2019;1(3):239–50. doi:10.31635/ccschem.019.20190010.
97. Boussuges A, Rives S, Marlinge M, Chaumet G, Vallée N, Guieu R, et al. Hyperoxia during exercise: impact on adenosine plasma levels and hemodynamic data. *Front Physiol.* 2020;11:252. doi:10.3389/fphys.2020.00097.
98. Sethumadhavan S, Silva M, Philbrook P, Nguyen T, Hatfield SM, Ohta A, et al. Hypoxia and hypoxia-inducible factor (HIF) downregulate antigen-presenting MHC class I molecules limiting tumor cell recognition by T cells. Simos G, editor. *PLoS One.* 2017;12(11):e0187314.
99. Hatfield SM, Kjaergaard J, Lukashev D, Belikoff B, Schreiber TH, Sethumadhavan S, et al. Systemic oxygenation weakens the hypoxia and hypoxia inducible factor 1 $\alpha$ -dependent and extracellular adenosine-mediated tumor protection. *J Mol Med Berl Ger.* 2014;92(12):1283–92. doi:10.1007/s00109-014-1189-3.
100. Liu X, Qin H, Zhang L, Jia C, Chao Z, Qin X, et al. Hyperoxia induces glucose metabolism reprogramming and intracellular acidification by suppressing MYC/MCT1 axis in lung cancer. *Redox Biol.* 2023;61:102647. doi:10.1016/j.redox.2023.102647.
101. Zhang L, Ke J, Min S, Wu N, Liu F, Qu Z, et al. Hyperbaric oxygen therapy represses the Warburg effect and epithelial-mesenchymal transition in hypoxic NSCLC cells via the HIF-

- 1a/PFKP axis. *Front Oncol.* 2021;11:691762. doi:10.3389/fonc.2021.691762.
102. Wang Y, Qi Y, Wei X, Chen S, Jia N, Zhou Q, et al. Hyperbaric oxygen rescues lung cancer cells from chemical hypoxia-induced low differentiation and apoptosis resistance. *Exp Lung Res.* 2018;44(8–9):417–23. doi:10.1080/01902148.2019.1571124.
103. Stellingwerff T, Leblanc PJ, Hollidge MG, Heigenhauser GJF, Spriet LL. Hyperoxia decreases muscle glycogenolysis, lactate production, and lactate efflux during steady-state exercise. *Am J Physiol Endocrinol Metab.* 2006;290(6):E1180–90. doi:10.1152/ajpendo.00499.2005.
104. Wang X, Ye N, Xu C, Xiao C, Zhang Z, Deng Q, et al. Hyperbaric oxygen regulates tumor mechanics and augments Abraxane and gemcitabine antitumor effects against pancreatic ductal adenocarcinoma by inhibiting cancer-associated fibroblasts. *Nano Today.* 2022;44:101458. doi:10.1016/j.nantod.2022.101458.
105. Liu X, Ye N, Liu S, Guan J, Deng Q, Zhang Z, et al. Hyperbaric oxygen boosts PD-1 antibody delivery and T cell infiltration for augmented immune responses against solid tumors. *Adv Sci WeinH Baden-Wurtt Ger.* 2021;8(15):e2100233.
106. Wu X, Zhu Y, Huang W, Li J, Zhang B, Li Z, et al. Hyperbaric oxygen potentiates doxil antitumor efficacy by promoting tumor penetration and sensitizing cancer cells. *Adv Sci.* 2018;5(8):1700859. doi:10.1002/advs.201700859.
107. Li ZH, Zhang X, Wu FG. Hyperbaric oxygen-facilitated cancer treatment: a minireview. *Adv NanoBiomed Res.* 2024;4(6):2300162. doi:10.1002/anbr.202300162.
108. Gawdi R, Yrastorza J, Cooper JS. Hyperbaric oxygen therapy contraindications. *StatPearls. Treasure Island (FL): StatPearls Publishing; 2025 [cited 2025 May 13]. Available from: <http://www.ncbi.nlm.nih.gov/books/NBK557661/>.*
109. Kim DW, Kim SW, Camidge DR, Shu CA, Marrone KA, Le X, et al. CD73 inhibitor oleclumab plus osimertinib in previously treated patients with advanced T790M-negative EGFR-mutated NSCLC: a brief report. *J Thorac Oncol.* 2023;18(5):650–6. doi:10.1016/j.jtho.2022.12.021.
110. Bendell JC, LoRusso P, Overman MJ, Noonan AM, Kim DW, Strickler J, et al. Safety and efficacy of the anti-CD73 monoclonal antibody (mAb) oleclumab ± durvalumab in patients (pts) with advanced colorectal cancer (CRC), pancreatic ductal adenocarcinoma (PDAC), or EGFR-mutant non-small cell lung cancer (EGFRm NSCLC). *J Clin Oncol.* 2021;39(15\_suppl):9047–7. doi:10.1200/jco.2018.36.15\_suppl.4123.
111. Hay CM, Sult E, Huang Q, Mulgrew K, Fuhrmann SR, McGlinchey KA, et al. Targeting CD73 in the tumor microenvironment with MEDI9447. *OncoImmunology.* 2016;5(8):e1208875. doi:10.1080/2162402x.2016.1208875.
112. Ghalamfarsa G, Rastegari A, Atyabi F, Hassannia H, Hojjat-Farsangi M, Ghanbari A, et al. Anti-angiogenic effects of CD73-specific siRNA-loaded nanoparticles in breast cancer-bearing mice. *J Cell Physiol.* 2018;233(10):7165–77. doi:10.1002/jcp.26743.
113. Tolcher AW, Gordon M, Mahoney KM, Seto A, Zavodovskaya M, Hsueh CH, et al. Phase 1 first-in-human study of dalutrafusp alfa, an anti-CD73-TGF- $\beta$ -trap bifunctional antibody, in patients with advanced solid tumors. *J Immunother Cancer.* 2023;11(2):e005267. doi:10.1136/jitc-2022-005267.
114. Anderson AE, Parashar K, Jin K, Clor J, Stagnaro CE, Vani U, et al. Characterization of AB598, a CD39 enzymatic inhibitory antibody for the treatment of solid tumors. *Mol Cancer Ther.* 2024;23(10):1471–82.
115. Spatola BN, Lerner AG, Wong C, Dela Cruz T, Welch M, Fung W, et al. Fully human anti-CD39 antibody potently inhibits ATPase activity in cancer cells via uncompetitive allosteric mechanism. *mAbs.* 2020;12(1):1838036.
116. Zhu H, Li X, Deng Q, Zhang Y, Wang H, He X, et al. Enhanced anti-ovarian cancer efficacy with MSLN-chimeric antigen receptor T cells secreting anti-CD39 antibody. *J Clin Oncol.* 2024;42(16\_suppl):e17553. doi:10.1200/jco.2024.42.16\_suppl.e17553.
117. Grigalavicius M, Ezzatpanah S, Papakiriakou A, Raabe TTH, Yannakopoulou K, Theodossiou TA. 5-ALA is a potent lactate dehydrogenase inhibitor but not a substrate: implications for cell glycolysis and new avenues in 5-ALA-mediated anticancer action. *Cancers.* 2022;14(16):4003.
118. Yang C, Xing S, Wei X, Lu J, Zhao G, Ma X, et al. 12-O-deacetylphomoxanthone A inhibits ovarian tumor growth and metastasis by downregulating PDK4. *Biomed Pharmacother Biomedicine Pharmacother.* 2024;175(3):116736. doi:10.1016/j.biopha.2024.116736.
119. Yang G, Li H, Yin J, Yao L, Yang J, Tang J, et al. Alleviating tumor hypoxia and immunosuppression via sononeoperfusion: a new ally for potentiating anti-PD-L1 blockade of solid tumor. *Ultrason Sonochem.* 2025;112(8):107115. doi:10.1016/j.ultrasonch.2024.107115.
120. Tambe Y, Terado T, Kim CJ, Mukaisho KI, Yoshida S, Sugihara H, et al. Antitumor activity of potent pyruvate dehydrogenase kinase 4 inhibitors from plants in pancreatic cancer. *Mol Carcinog.* 2019;58(10):1726–37. doi:10.1002/mc.23045.
121. Gao F, Tang Y, Liu WL, Zou MZ, Huang C, Liu CJ, et al. Intra/Extracellular lactic acid exhaustion for synergistic metabolic therapy and immunotherapy of tumors. *Adv Mater Deerfield Beach Fla.* 2019;31(51):e1904639. doi:10.1002/adma.201904639.
122. Franczak MA, Krol O, Harasim G, Jedrzejewska A, Zaffaroni N, Granchi C, et al. Metabolic effects of new glucose transporter (GLUT-1) and lactate dehydrogenase-A (LDH-A) inhibitors against chemoresistant malignant mesothelioma. *Int J Mol Sci.* 2023;24(9):7771. doi:10.3390/ijms24097771.
123. Gayatri MB, Kancha RK, Patchva D, Velugonda N, Gundeti S, Reddy ABM. Metformin exerts antileukemic effects by modulating lactate metabolism and overcomes imatinib resistance in chronic myelogenous leukemia. *FEBS J.* 2023;290(18):4480–95. doi:10.1111/febs.16818.
124. Masjedi A, Ahmadi A, Ghani S, Malakotikhah F, Nabi Afjadi M, Irandoust M, et al. Silencing adenosine A2a receptor enhances dendritic cell-based cancer immunotherapy. *Nanomedicine Nanotechnol Biol Med.* 2020;29:102240. doi:10.1016/j.nano.2023.102690.
125. Alobaidi B, Hashimi SM, Alqosaibi AI, AlQurashi N, Alhazmi S. Targeting the monocarboxylate transporter MCT2 and lactate dehydrogenase A LDHA in cancer cells with FX-11 and AR-C155858 inhibitors. *Eur Rev Med Pharmacol Sci.* 2023;27(14):6605–17.
126. Aphale R, Shah SM. A randomised clinical trial to compare the efficacy of hyperbaric oxygen therapy with neoadjuvant chemotherapy with neoadjuvant chemotherapy alone for

- carcinoma breast: a pilot study. *Indian J Surg.* 2021;83(S2):511–5. doi:10.1007/s12262-020-02601-4.
127. Mink Van Der Molen DR, Batenburg MCT, Maarse W, Van Den Bongard DHJG, Doeksen A, De Lange MY, et al. Hyperbaric oxygen therapy and late local toxic effects in patients with irradiated breast cancer: a randomized clinical trial. *JAMA Oncol.* 2024;10(4):464. doi:10.1001/jamaoncol.2023.6776.
  128. Hu Y, Sarkar A, Song K, Michael S, Hook M, Wang R, et al. Selective refueling of CAR T cells using ADA1 and CD26 boosts antitumor immunity. *Cell Rep Med.* [cited 2025 May 13]. Available from: [https://www.cell.com/cell-reports-medicine/abstract/S2666-3791\(24\)00199-X](https://www.cell.com/cell-reports-medicine/abstract/S2666-3791(24)00199-X).
  129. Qu Y, Dunn ZS, Chen X, MacMullan M, Cinay G, Wang HY, et al. Adenosine deaminase 1 overexpression enhances the antitumor efficacy of chimeric antigen receptor-engineered T cells. *Hum Gene Ther.* 2022;33(5–6):223–36. doi:10.1089/hum.2021.050.
  130. Zeng W, Liu P, Pan W, Singh SR, Wei Y. Hypoxia and hypoxia inducible factors in tumor metabolism. *Cancer Lett.* 2015;356(2):263–267. doi:10.1016/j.canlet.2014.01.032.
  131. Eales KL, Hollinshead KER, Tennant DA. Hypoxia and metabolic adaptation of cancer cells. *Oncogenesis.* 2016;5(1):e190. doi:10.1038/oncsis.2015.50.
  132. Kumar V, Gabrilovich DI. Hypoxia-inducible factors in regulation of immune responses in tumour microenvironment. *Immunology.* 2014;143(4):512–9. doi:10.1111/imm.12380.
  133. Corcoran SE, O'Neill LAJ. HIF1 $\alpha$  and metabolic reprogramming in inflammation. *J Clin Invest.* 2016;126(10):3699–707. doi:10.1172/jci84431.